

Megavoltage X-ray Dose Enhancement with Gold Nanoparticles in Tumor Bearing Mice

Sayyed Hossein Mousavie Anijdan^{1*}, Seyyed Rabi Mahdavi², Alireza Shirazi²,
Mohammad Ali Zarrinfard³, Jamshid Hajati⁴

1. Department of Medical Physics, Babol University of Medical Sciences, Babol, Iran.
2. Department of Medical Physics and Engineering, School of Medicine, Tehran University of Medical Sciences, Tehran, Iran.
3. Department of Medical Nanotechnology, Faculty of Advanced Medical Sciences, Tehran University of Medical Sciences, Tehran, Iran.
4. Department of Immunology, Faculty of Medicine, Tehran University of Medical Sciences, Tehran, Iran.

Submitted 13 July 2013; Accepted 4 Aug 2013

One of the applications of gold nanoparticles (GNPs) in medicine is radiation dose-enhancing effect. Although there are many simulations, *in vitro* and *in vivo* evidence that GNPs can enhance significantly the radiation dose effect of orthovoltage beams. These beams compared with megavoltage (MV) beams, have limited applications in radiotherapy. In order to evaluate GNPs radiosensitization performance with MV beams *in-vivo*, we used two most clinically used X-ray beams (6 and 18 MV) with the dose of 20 Gy for each mouse. Intratumoral injection of 50 nm GNPs with the concentration of 5 mg ml⁻¹ was applied to melanoma tumor growing in the left leg of 7 to 8 mice in 4 control and treatment groups of C57BL/6 mice. Albeit, using 10 cm plexiglass jig phantom in the beam path improved the radiation - treatments, the statistical differences between groups were not significant. According to the results, it is concluded that mice can be treated with smaller tumors and higher concentrations of GNPs in MV radiation beams.

Key words: Gold nanoparticles (GNPs), radiation dose enhancement, tumor, megavoltage X-ray

Radiotherapy is an important method in cancer treatment, but it can be held responsible for biological damages as radiations can also induce a decrease in normal tissues. Improvements in this cancer therapeutic method have begun in the past decades by performing intensity modulated radiation therapy (IMRT), multileaf collimators (MLCs) and stereotactic radiosurgery. Radiosen-

sitization of tumors with safety media is an alternative method to improve the discrimination between tumors and normal tissues (1-3). Initially, this technique was known as X-ray phototherapy with iodine and gadolinium contrast media (4-5). Then it was introduced as contrast-enhanced radiation therapy or CERT (6-7) but in numerous papers, this method was mentioned as gold

* Corresponding author: Department of Medical Physics, Babol University of Medical Sciences, Babol, Iran. Email mousavi@razi.tums.ac.ir

nanoparticle radiation therapy or GNRT (8-9). In recent years, this method has been of an increasingly interest by the use of gold nanoparticles radiosensitize contrast media due to its high atomic number and bio-compatibility (8). Through the progress in nanotechnology, the synthesis of various gold nanostructures such as spherical gold nanoparticles (GNPs), gold nanorods (GNRs), gold nanoshells (GNSs) and gold nanocages (GNCs) in cancer therapeutics have been made (2). Gold nanostructures can be applied as radiation sensitizers, anticancer drug enhancers, heat generators and also effective drug carriers.

The first step of this study was a series of computer simulations (10). The Monte Carlo method was described in several studies in the last years. These studies have all shown the radiosensitization of GNPs, especially with kilovoltage radiation exposures (11-14). But recent studies have also shown the radiosensitization effect at megavoltage beams (15-17). This effect can be understood on the basis of Local Effect Model (LEM) at nanometer scale.

Many other *in vitro* studies have also demonstrated dose enhancement with different cells at low voltage X-rays (18-22). Jain et al. (2010), for instance, performed clinically relevant MV X-ray with 2 nm GNPs and obtained comparably the same effect at kilovoltage (23). A recent *in vitro* study has shown radiation sensitizer enhancement ratios (SERs) of 1.29 and 1.16 at using 6 MV and 15 MV, respectively (24). Another recent study has indicated the relative biological effectiveness of proton beam on prostate tumor cells, approximately 15%-20% radiosensitization when cells contain internalized 44 nm of GNPs (25).

Animal studies have also shown that gold microspheres or nanoparticles can enhance radiobiological effect of radiation dose (18, 26-29). In some of these studies with the enhanced permeation and retention (EPR) effect, nanoparticles can accumulate in tumor through

passive targeting (30). Hainfeld et al. were the first researchers who successfully demonstrated long time animal study with relatively high concentration 2 nm GNPs passively (26). Apoptotic potential and dose-enhancing effect of clinical electron beams were investigated on B16F10 melanoma tumor-bearing mice (27). Therefore, in this study, we investigated the dose-enhancing effect of gold nanoparticles in combination with single-dose clinical 6 and 18 MV photon beams on B16F10 melanoma tumor-bearing mice.

Materials and Methods

Cells and mice

One T₂₅ flask of murine B16F10 melanoma cells was obtained from National Cell Bank of Pasteur Institute of Iran (NCBI, C542). These cells were cultured in RPMI 1640 (Gibco G002909-2037) medium supplemented with 2 nM L-glutamine, Penn/ Strep (100 U ml⁻¹ Penicillin; 100 µg ml⁻¹ Streptomycin) and fetal bovine serum (Gibco A15111-0229) at 37 °C in a pressure of 5% CO₂. After two or three subcultures, the cells were prepared for injection into the mice. Female C57BL/6 mice (8-10 weeks old) were also obtained from the Karaj Production and Research Center Laboratory of Animal Science Department of the Pasteur Institute of Iran. All the experiments performed on mice were also approved by the Animal Care Committee of Tehran University of Medical Sciences. These C57BL/6 mice were planted subcutaneously (s.c.) in the thigh with 150000 murine isogenic melanoma B16F10 cultured cells suspended in PBS in a total volume of 40-50 µl. After 2 weeks, the tumors volume reached about 400-600 mm³ (Fig. 1).

Tumor volume was estimated by measuring three diameter of mass and calculated with orthogonal diameters equation (31). One group of tumor bearing mice (including eight mice) had direct intratumoral injection with 100-200 µl of 5 mg ml⁻¹ GNPs. All mice were monitored for the

growth of tumor and survival during the experiments.

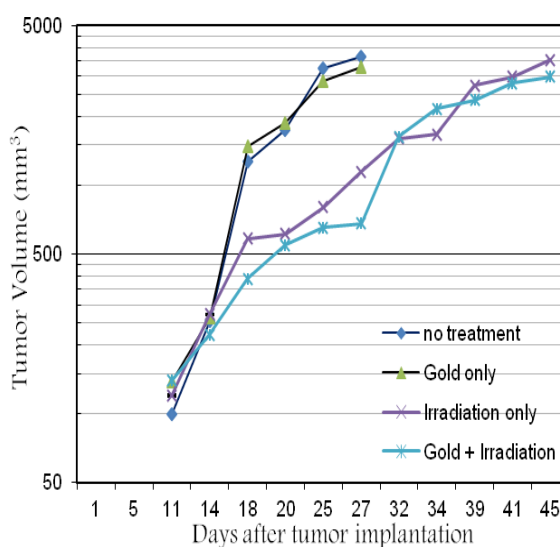


Fig. 1. Average tumor volume at different groups of mice irradiated with 18 MV after: no treatment (diamonds, PBS); gold only (triangles); irradiation only (cross); intratumoral gold injection followed by irradiation (stars).

Gold nanoparticles and biodistribution

50 ± 2 nm gold nanoparticles (Plasmachem, Germany) were injected intratumorally at several sites. The volume level of GNPs injected to mice was in accordance with their accumulation in tumor proposed by Hainfeld et al. (26, 28). The GNPs concentration was about 7 mg ml^{-1} in the tumor. Digital radiographic images of the mice were used to identify intratumoral distribution of GNPs.

Characteristics of irradiation

The mice were anesthetized with Xiluzin and Ketamin intraperitoneally. About 10 mm in diameter of the region of mice leg containing the tumor was irradiated with 6 and 18 MV X-rays through Varian 2100 C/D linear accelerator (LINAC) approximately 30 min post GNPs injection. The output rate was 300 MU/min and the total delivered dose was 20 Gy.

It was not possible to perform this high single fraction dose to this depth tumor with LINAC for each mouse separately. Therefore a special phantom was designed that consisted of two separate slabs. The upper slab with the thickness of

10 cm plexiglass is thick enough to produce transient charged particle equilibrium for 6 and 18 MV X-ray beams. The lower slab was made from wax with eight places for eight tumor bearing legs of mice (Fig. 2). Radiation was applied to an $8.5 \text{ cm} \times 20 \text{ cm}$ field size at 108 cm source-to-surface distance (SSD). The total 20 Gy dose was delivered with five 576 MU's separated very short intervals. The time within each fraction was at least 5 minute.



Fig. 2. Set-ups of mice irradiation with plexiglass phantom and wax formed slab.

Results

B16F10 melanoma cells were cultured and grown subcutaneously in the left legs of the C57BL/6 mice. The growth of the tumor and mice survival

were also considered indices for the different treatments groups. Figure 1 shows the average tumor volumes of the different groups of mice irradiated with 18 MV. As shown in this figure, the rate of tumor size in control tumor bearing mice (no treatment and gold only) was different with the group receiving radiation only and the mice with GNPs and radiation. The average tumor volumes after various treatment modalities were meaningfully different between control groups and treatment groups. But this difference is not significant between irradiation only and gold with irradiation groups ($P>0.05$). It is depicted that in the first month after irradiation, tumor size control in gold with irradiation groups was better than irradiation groups only.

Figure 3 shows the tumor volume doubling time for three groups. Growth speed of tumor is identified from the slope of these lines as 0.381, 0.246 and 0.147 for the three groups 2-4. The volume doubling time were obtained 44, 76 and 113 hr for groups 2-4, respectively.

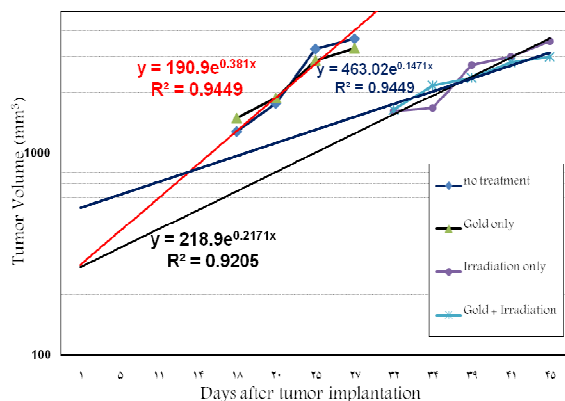


Fig. 3. Tumor volume doubling time at different groups of mice irradiated with 18 MV.

Kaplan-Meier survival graphs (Fig. 4) have also shown the differences between groups. The GNPs without radiation could not stop the growth of the tumor and increased the survival of the mice. All mice receiving either no radiation or GNPs without radiation died during the span of one month.

Discussion

In this study, *in vivo* GNPs radiosensitization was evaluated in radiotherapy with clinical megavoltage X-ray beams. We used the same geometrical set-ups utilized in the separate simulation and dosimetric study (10). DEF between 8-10% was obtained in deep tumor-like insert phantom (10 cm depth) that contained GNPs and irradiated with 18 MV.

Survival of four groups: Survival proportions

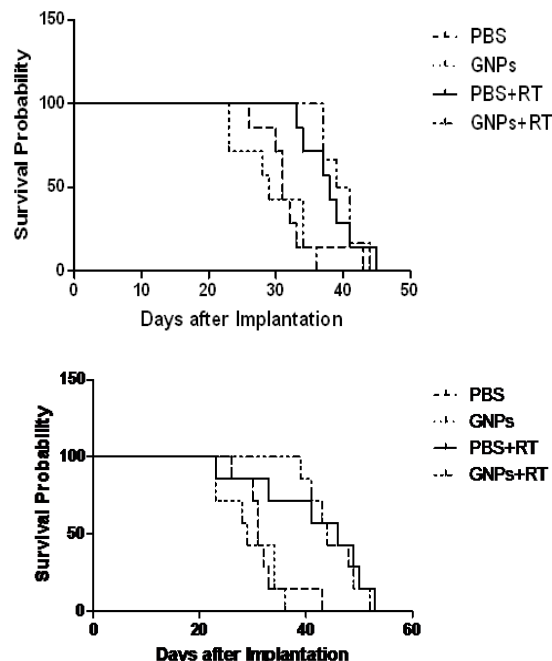


Fig. 4. Kaplan-Meier survival graph of mice with different treatments, with 6 MV (up) and 18 MV (down).

It appears that the size of tumor is a very important parameter in radiotherapy. This study is one of the novel works that deals with high energy X-ray beams. It is revealed that tumor size and its response to the various treatments determine the shape of cell survival probability. We carried out treatments with about 500 mm³ tumor size while Hainfeld and Chang performed their *in vivo* studies with very small tumor sizes (100 mm³ approximately) (26-27). Another motive was GNPs desired concentration in the tumor. Thus, tumor size was only controlled in the first month after irradiation with 18 MV (Fig. 1). This effect occurred much less

in 6 MV. As seen in figure 3, the tumor size doubling time shows the topic. Because GNPs were injected intratumorally, the issue of the high liver uptake was not important for GNPs with 50 nm (26). Results reported from the MC simulation studies have shown that nanoparticles greater than 50 nm provide a more significant dose enhancement (12). An *in vitro* study by Chithrani et al. shows that cellular uptake is maximized when the size of nanoparticles is 50 nm (19). However, direct intra-tumoral injection has the potential disadvantage of non-uniform distribution, but in this study the GNPs were injected in several directions and sites in the tumor (18).

Jain et al. conducted experimental studies with 2 nm gold nanoparticles and observed a comparable dose enhancement in megavoltage similar to the kilovoltage CERT (23). They concluded that the increased X-ray absorption was not the main mechanism of physical dose enhancement. Also, it can be considered that there is a discrepancy between the physical dose enhancement in X-ray energy deposition and biologically observed dose enhancement in cell killing with different GNP sizes that are explained below (32). Usually, MC simulations consider GNP as a single particle and calculate the deposited energy with the radiation interaction. If the sizes of the GNPs are small (e.g. 2 nm), the interaction frequency and dose enhancement will be small and vice-versa. Also the results of Leung et al. show that GNP with greater sizes increases the generation of secondary electrons. However, inside the cells, small-sized GNPs form a single large cluster within the vesicles with a size of about 300-500 nm (12).

A new theoretical approach in calculating radiosensitization is the microscopic dose enhancement that is related to the deposited dose on the walls of tumor blood vessels (15, 33-34). Berbeco et al. revealed a 1.7 endothelial dose enhancement factor (EDEF) after the application of 30 mg ml⁻¹ intravascular GNPs concentration for 6

MV beam at the depth of 20 cm (17). Furthermore they reported high EDEF values of 1.2 to 4.4 (24-340% dose increase) that happen in the vicinity of the nanoparticles. These findings also help us to explain the DEF discrepancy between our results and Robar and Cho (5, 8). Contribution of 10 cm slab phantom above the GNPs container should be considered to produce low energy X-rays and scattered radiation from 6 and 18 MV primary incident photons which cause dose enhancement within the tumor immediately after GNPs through appropriate interaction mechanisms.

Our study has shown that local dose enhancement can be achieved by the GNPs with 6 and 18 MV photon beams at the depth of 10 cm. It also shows that the method of nanoparticles distribution can effectively change the DEF. In spite of all relevant dosimetric and simulation results that confirm dose enhancement, more research is suggested to be done for more biological clue. To achieve biological evidence, additional *in vivo* experimental set ups similar to this study should be conducted to investigate as to whether gold nanoparticles have the potential to be used as a radiosensitizer in radiation therapy of tumors.

Acknowledgments

This research was supported by a grant from the Tehran University of Medical Sciences 89-04-30-11442.

References

1. Samarasinghe RM, Kanwar RK, Kanwar JR. The role of nanomedicine in cell Based therapeutics in cancer and inflammation. *Int J Mol Cell Med* 2012;1:133-44.
2. Jelveh S, Chithrani DB. Gold Nanostructures as a Platform for Combinational Therapy in Future Cancer Therapeutics. *Cancers* 2011;3:1081-110.
3. Butterworth KT, McMahon SJ, Currell FJ, et al. Physical basis and biological mechanisms of gold nanoparticle radiosensitization. *Nanoscale* 2012;4:4830-8.
4. Mesa AV, Norman A, Solberg TD, et al. Dose distributions using kilovoltage x-rays and dose enhancement from iodine

contrast agents. *Phys Med Biol* 1999;44:1955-68.

5. Robar JL, Riccio SA, Martin MA. Tumour dose enhancement using modified megavoltage photon beams and contrast media. *Phys Med Biol* 2002;47:2433-49.

6. Verhaegen F, Reniers B, Deblois F, et al. Dosimetric and microdosimetric study of contrast-enhanced radiotherapy with kilovolt x-rays. *Phys Med Biol* 2005;50:3555-69.

7. Morris KN, Weil MD, Malzbender R. Radiochromic film dosimetry of contrast-enhanced radiotherapy (CERT). *Phys Med Biol* 2006;51:5915-25.

8. Cho SH. Estimation of tumour dose enhancement due to gold nanoparticles during typical radiation treatments: a preliminary Monte Carlo study. *Phys Med Biol* 2005;50:N163-73.

9. Cho SH, Jones BL, Krishnan S. The dosimetric feasibility of gold nanoparticle-aided radiation therapy (GNRT) via brachytherapy using low-energy gamma-/x-ray sources. *Phys Med Biol* 2009;54:4889-905.

10. Mousavie Anijdan SH, Shirazi A, Mahdavi SR, et al. Megavoltage dose enhancement of gold nanoparticles for different geometric set-ups: Measurements and Monte Carlo simulation. *Ir J Radiat Res* 2012;10:183-86.

11. Zhang SX, Gao J, Buchholz TA, et al. Quantifying tumor-selective radiation dose enhancements using gold nanoparticles: a monte carlo simulation study. *Biomed Microdevices* 2009;11:925-33.

12. Leung MK, Chow JC, Chithrani BD, et al. Irradiation of gold nanoparticles by x-rays: Monte Carlo simulation of dose enhancements and the spatial properties of the secondary electrons production. *Med Phys* 2011;38:624-31.

13. Lechtman E, Chattopadhyay N, Cai Z, et al. Implications on clinical scenario of gold nanoparticle radiosensitization in regards to photon energy, nanoparticle size, concentration and location. *Phys Med Biol* 2011;56:4631-47.

14. Lechtman E, Mashouf S, Chattopadhyay N, et al. A Monte Carlo-based model of gold nanoparticle radiosensitization accounting for increased radiobiological effectiveness. *Phys Med Biol* 2013;58:3075-87.

15. McMahon SJ, Hyland WB, Muir MF, et al. Biological consequences of nanoscale energy deposition near irradiated heavy atom nanoparticles. *Sci Rep* 2011;1:1-9.

16. McMahon SJ, Hyland WB, Muir MF, et al. Nanodosimetric effects of gold nanoparticles in megavoltage radiation therapy.

Radiother Oncol 2011;100:412-6.

17. Berbeco RI, Ngwa W, Makrigiorgos GM. Localized dose enhancement to tumor blood vessel endothelial cells via megavoltage X-rays and targeted gold nanoparticles: new potential for external beam radiotherapy. *Int J Radiat Oncol Biol Phys* 2011;81:270-6.

18. Herold DM, Das JJ, Stobbe CC, et al. Gold microspheres: a selective technique for producing biologically effective dose enhancement. *Int J Radiat Biol* 2000;76:1357-64.

19. Chithrani BD, Ghazani AA, Chan WC. Determining the size and shape dependence of gold nanoparticle uptake into mammalian cells. *Nano Lett* 2006;6:662-8.

20. Zhang X, Xing JZ, Chen J, et al. Enhanced radiation sensitivity in prostate cancer by gold-nanoparticles. *Clin Invest Med* 2008;31:E160-7.

21. Roa W, Zhang X, Guo L, et al. Gold nanoparticle sensitize radiotherapy of prostate cancer cells by regulation of the cell cycle. *Nanotechnology* 2009;20:375101-10.

22. Chithrani DB, Jelveh S, Jalali F, et al. Gold nanoparticles as radiation sensitizers in cancer therapy. *Radiat Res* 2010;173:719-28.

23. Jain S, Coulter J, Butterworth K, et al. Gold nanoparticles as sensitizers for radiation therapy at clinically relevant megavoltage X-ray energies. *Genitourinary Cancers Symposium*; New York: ASCO University; 2010. p. 126.

24. Jain S, Coulter JA, Hounsell AR, et al. Cell-specific radiosensitization by gold nanoparticles at megavoltage radiation energies. *Int J Radiat Oncol Biol Phys* 2011;79:531-9.

25. Polf JC, Bronk LF, Driessen WH, et al. Enhanced relative biological effectiveness of proton radiotherapy in tumor cells with internalized gold nanoparticles. *Appl Phys Lett* 2011;98:193702-3.

26. Hainfeld JF, Slatkin DN, Smilowitz HM. The use of gold nanoparticles to enhance radiotherapy in mice. *Phys Med Biol* 2004;49:N309-15.

27. Chang MY, Shiau AL, Chen YH, et al. Increased apoptotic potential and dose-enhancing effect of gold nanoparticles in combination with single-dose clinical electron beams on tumor-bearing mice. *Cancer Sci* 2008;99:1479-84.

28. Hainfeld JF, Dilmanian FA, Zhong Z, et al. Gold nanoparticles enhance the radiation therapy of a murine squamous cell carcinoma. *Phys Med Biol* 2010;55:3045-59.

29. Zhang XD, Wu HY, Wu D, et al. Toxicologic effects of gold nanoparticles in vivo by different administration routes. *Int J Nanomedicine* 2010;5:771-81.
30. Maeda H. The enhanced permeability and retention (EPR) effect in tumor vasculature: the key role of tumor-selective macromolecular drug targeting. *Adv Enzyme Regul* 2001;41:189-207.
31. Venslauskas MB, D. B. Irregular solid tumor growth. Estimation of electrochemotherapy efficacy. *Acta medica Lituanica* 2003;10:28-32.
32. Mesbahi A. A review on gold nanoparticles radiosensitization effect in radiation therapy of cancer. *Reports of Practical Oncology & Radiotherapy* 2010;15:176-80.
33. Ngwa W, Makrigiorgos GM, Berbeco RI. Applying gold nanoparticles as tumor-vascular disrupting agents during brachytherapy: estimation of endothelial dose enhancement. *Phys Med Biol* 2010;55:6533-48.
34. Jones BL, Krishnan S, Cho SH. Estimation of microscopic dose enhancement factor around gold nanoparticles by Monte Carlo calculations. *Med Phys* 2010;37:3809-16.