IJMCM
Autumn 2012, Vol 1, No 4

Original Article

Rab11 is required for maintenance of cell shape via βPS integrin mediated cell adhesion in *Drosophila*

Tanmay Bhuin, Jagat Kumar Roy*

Cytogenetics Laboratory, Department of Zoology, Banaras Hindu University, Varanasi 221 005, India.

Submmited 13 January 2013; Accepted 8March 2013

In eukaryotes, vesicle trafficking is regulated by the small monomeric GTPases of the Rab protein family. Rab11, (a subfamily of the *Ypt/Rab* gene family) an evolutionarily conserved, ubiquitously expressed subfamily of small monomeric Rab GTPases, has been implicated in regulating vesicular trafficking through the recycling of endosomal compartment. In an earlier communication, we have shown that Rab11 is required for cell adhesion, maintenance of cell shape and actin-cytoskeleton organization during *Drosophila* wing development. Here, we report that Rab11 is required for the maintenance of cell shape via βPS integrin mediated cell adhesion. When Rab11 is over-expressed or activity of Rab11 is reduced via a double-stranded RNAi line, the cuticular preparation of the embryos show dorsal open phenotypes. Immuno-fluorescence and immuno-histochemical analyses on embryos in the same genetic backgrounds also show altered localization of βPS integrins at the adhesion site of leading edge and amnioserosa cells during the dorsal closure stages of embryogenesis. The cellular morphology (cell shape) of the lateral epidermal cells are also affected.

Key words: Amnioserosa, cellular morphology, cuticle, dorsal closure, *Drosophila*, Rab11

Intra-cellular vesicle transport is a complex process mediated by a diverse array of proteins. A large group of small monomeric GTP-binding proteins, Rabs, forming a major class of Ras superfamily (1), are essential component of this process. These proteins are associated with various exocytic and endocytic organelles, as well as vesicles that are coupled with these compartments. Rab proteins regulate vesicular trafficking pathways, behaving as membrane-associated molecular switches (2). *Rab11* is an evolutionary conserved ubiquitously expressed subfamily of the *Ypt/Rab* gene family and is associated with recycling of endosomes (RE). Multiple roles of *Rab11* have been characterized in *Drosophila* during development, differentiation and signaling pathways. Rab11 endosomes are involved in the regulation of vesicle exocytosis and membrane growth during cellularization, a special form of embryonic cleavage in *Drosophila* (3) and is also essential for the invagination of furrows during *Drosophila* embryonic cleavage (4). Previous studies have shown that Rab11 is required for the process of myoblast fusion (5) and in the

* Corresponding author: Cytogenetics Laboratory, Department of Zoology, Banaras Hindu University, Varanasi 221 005, India.
E-mail: jkroy@bhu.ac.in

[Downloaded from ijmcmed.org on 2026-10-15]

development of nervous system during *Drosophila* embryogenesis (6).

Dorsal closure (DC) in *Drosophila* is an important morphogenetic event that takes place during stages 13-15 of embryogenesis. It covers the dorsal hole and establishes the dorsal ectoderm by stretching the lateral epithelial cells over the amnioserosa (7). This process relies strictly upon coordinated changes in cell shape by cell elongation, without cell recruitment or proliferation (8). In embryos, Rab11 localizes at significant levels both in the lateral epidermal cells and in the amnioserosa during dorsal closure. It has been shown that loss of Rab11 function leads to improper dorsal closure in the developing *Drosophila* embryo (9).

Integrins are one of the best characterized and major cell surface receptors which mediate cellular connections between the ligands in the extracellular matrix to the actin cytoskeleton. They are heterodimeric protein complexes composed of an α and a β subunit. They are involved in the attachment of cells with the extra cellular matrix and also for the transmission of various signals from the extracellular environment into various signaling pathways in cells (10, 11).

Drosophila integrins serve as prototype for the genetic studies of cell adhesion proteins during development. Various functions of integrins in *Drosophila* development have been extensively studied like maintaining the close apposition of wing surfaces during wing morphogenesis (12), attachment of embryonic muscle to epidermis and mediating the connections between lateral surface of amnioserosa cells and leading edge (LE) cells during embryonic dorsal closure (13). Evidences point to the fact that integrin-extracellular matrix interactions occur between the lateral surfaces of the amnioserosa cells and the LE cells that effectively mediates cell-cell adhesion. It has been seen that *Drosophila* embryos lacking βPS integrin have a hole in the dorsal epidermis due to the failure of proper dorsal closure (13). In a recent communication, we have shown that Rab11 is involved in the trafficking of βPS integrins during *Drosophila* wing development. The studies have also shown that alterations of Rab11 functions results in the failure of βPS integrins to reach the focal adhesions (14). Here, we report that *Rab11* affects the localization of βPS integrins from the adhesion site of leading edge and amnioserosa cells during the dorsal closure stages of embryogenesis and also changes cellular morphology/cell shape of the lateral epidermal (LAE) cells.

Materials and Methods

Fly stocks, genetics and lethality assay

The following stocks were used: *UAS-Rab11*^{RNAi} (15) (gift from D Ready), *UAS-Rab11*^{CA} (16) (a gift of M. Gonzalez-Gaitan). Other stocks were from Bloomington Stock Centre. In order to replace *CyO* balancers in *Rab11*^{CA} allele, reporter lacZ fused with the *CyO*, *CyO-ftz-lacZ*, balancer was introduced.

To determine the phenotypes generated by over-expressing *Rab11*^{CA} or *Rab11*^{RNAi} in epithelial tissue, the homozygous *GAL4* line of *prd-GAL4* was crossed to heterozygous *UAS-Rab11*^{CA}*/CyO-ftz-lacZ* or homozygous *UAS-Rab11*^{RNAi} flies so that, in the first case, 50% of the embryos would be driven by *GAL4* line, whereas 100% embryos would be driven in this manner in the case of the *Rab11*^{RNAi} line. Because of the presence of lacZ reporter in the balancer chromosome, the trans-heterozygotes with *prd-GAL4* or without *prd-GAL4* could easily be segregated by staining with anti-β-galactosidase antibody. Thus, unstained embryos obtained by blue balancer selection or embryos containing dsRNA driven by *prd-GAL4* were further processed for antibody (different markers) staining.

Homozygous virgins from *GAL4* line were crossed to homozygous *UAS-Rab11*^{RNAi} flies, and heterozygous constitutively active *UAS-Rab11*^{CA}/+

[Downloaded from ijmcmed.org on 2026-10-15]

(obtained after crossing with Oregon R flies) male flies and eggs/embryos from the different genotypes were collected on separate agar plates; the total number of embryos in each plate was counted. Unhatched embryos were counted after 25-30 hrs of egg laying (at 25°C), dechorionated and then observed for lethality. Unfertilized eggs were also counted and discarded while calculating embryonic lethality. In each case, at least 5000 embryos were counted. The percentage of embryonic lethality (EL) was calculated by using the following formulae: For homozygous *Rab11* allele ($Rab11^{RNAi}$): EL = Dead embryos X100/Total no. of fertilized eggs.

For heterozygous *Rab11* allele ($Rab11^{CA}$): EL = Dead embryos X 100/total no. of fertilized eggs X 2 [Total no. of fertilized eggs = total no. of eggs - no. of unfertilized eggs]

[Dead embryos = total no. of fertilized eggs-total no. of hatched embryos]

Immunostaining of embryos

Antibody staining was performed as described by Van Vactor and Kopczynski (17) with slight modifications. The primary antibodies used were as follows: mouse monoclonal anti-phosphotyrosine (Santa Crutz; 1:100), mouse anti-βPS integrin (CF6G11, DSHB; 1:10); alkaline-phosphatase-conjugated goat anti-rabbit IgG (Sigma; 1:1000). Secondary antibodies used were as follows: AF-488-conjugated goat anti-mouse, biotinylated goat anti-mouse IgG (Jackson Immunoresearch; 1:300). Biotinylated secondary antibodies were used in combination with the Vector Elite ABC kit (Vector Laboratories, Calif.) according to the manufacturer's instructions. For improved signal intensity and colour contrast, the staining was developed by using 0.5 mg/ml DAB (3, 3-diaminobenzidine) and 0.02% H_2O_2 (hydrogen peroxide); 8% nickel chloride was used for further enhancement of the regular DAB procedure. Fluorescent whole-mount embryos were mounted in 70% glycerol in phosphate-buffered saline (PBS) supplemented with 15% DABCO (1,4-diazabicyclo-[2.2.2]-octane) and images were taken on a BioRad MRC laser scanning confocal microscope, whereas horseradish peroxidase (HRP)-stained embryos were mounted in 70% glycerol in PBS and examined and photographed under bright-field optics. Images were processed with BioRad software and/or Adobe Photoshop.

Cuticle preparation from wild type and mutant embryos

Wild type embryos were allowed for a synchronised egg laying. After 4 hrs, the embryos were collected and allowed to develop for 17-22 hrs, so that the cuticle layer is secreted. The embryos were then dechorionised, fixed and devitalinized in the same way as it was done for immunostaining. However after the final wash with methanol the embryos were suspended in glycerol + acetic acid (1:4) solution at 65°C, overnight. They were then mounted in Hoyer's medium and again kept at 65°C, overnight. The cuticles were then visualized under a phase contrast microscope.

Mutant embryos were processed 48 hrs after collection from an overnight egg lying on an agar plate. The dead embryos were selected. They were then dechorionised and devitelinized in the same way. They were then suspended in glycerol and acetic acid (1:4) solution for overnight at 65°C, mounted in Hoyer's medium and kept at 65°C for overnight. The cuticles were then visualized under a phase contrast microscope.

Results and Discussion

Loss of Rab11 function leads to the disintegration of embryonic cuticles

Studies have shown that βPS integrin is essential for proper dorsal closure to occur (13). The strongly hypomorphic/nearest to null allele of *Rab11* show defects in dorsal closure and thus have defective cuticle (9). It has also been shown that Rab11 is involved in the trafficking of βPS integrin during *Drosophila* wing development (14).

[Downloaded from ijmcmed.org on 2026-10-15]

Therefore, it has been hypothesized that there might be a loss of βPS integrin in the leading edge epidermal cells in *Rab11* mutant backgrounds. To test this in a tissue specific manner and to study the phenotypic consequences, the constitutively-active ($Rab11^{CA}$) and the $Rab11^{RNAi}$ alleles were expressed using UAS-GAL4 system (18) in the epidermal stripes driven by *prd-GAL4* during dorsal closure stages of embryogenesis. Expression of these two *Rab11* transgenes with *prd-GAL4* causes embryonic to larval lethality (In case of $Rab11^{CA}$ alleles; 12% embryonic and rest larval lethality whereas in case of $Rab11^{RNAi}$ lines 10% embryonic and rest larval lethality were observed) which suggested further studies to analyse the epidermal development in these mutants. Cuticle preparations of these lethal embryos were carried out which showed presence of dorsal/anterior openings (Fig. 1B and 1C) as compared to the wild type (Fig. 1A) in conformity with the previous observation (9).

Inactivation of Rab11 function shows loss of βPS integrins from the leading edge epidermal cells

To observe whether there is any significant loss of βPS integrin from the leading edge epidermal cells in Rab11 mutant backgrounds, immunostaining of whole embryos of $Rab11^{CA}$ and $Rab11^{RNAi}$ driven by *prd-GAL4* was carried out using anti-βPS integrin-antibody. It was seen that there was a significant (80 embryos, n=200) loss of βPS integrin in the leading edge epidermal cells of both the alleles (Fig. 2B and 2C) when compared to the normal wild type embryos (Fig. 2A). However in the $Rab11^{RNAi}$ line aggregation of βPS integrin was also seen.

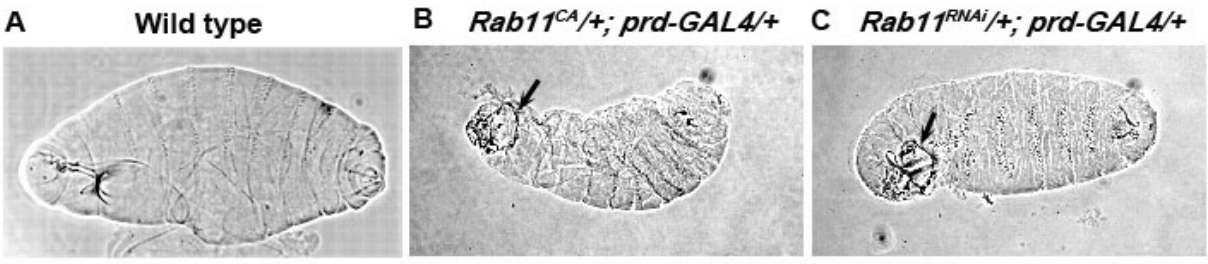

Fig 1. Cuticles of $Rab11^{CA}$ and $Rab11^{RNAi}$ mutant embryos displaying the presence of anterior/dorsal hole. Anterior is towards the left and the dorsal side is laterally placed in A, B and C. A, B and C show the cuticle of a wild type embryo and embryos from $Rab11^{CA}$ and $Rab11^{RNAi}$, respectively. The black arrows in B and C indicate the anterior/dorsal hole.

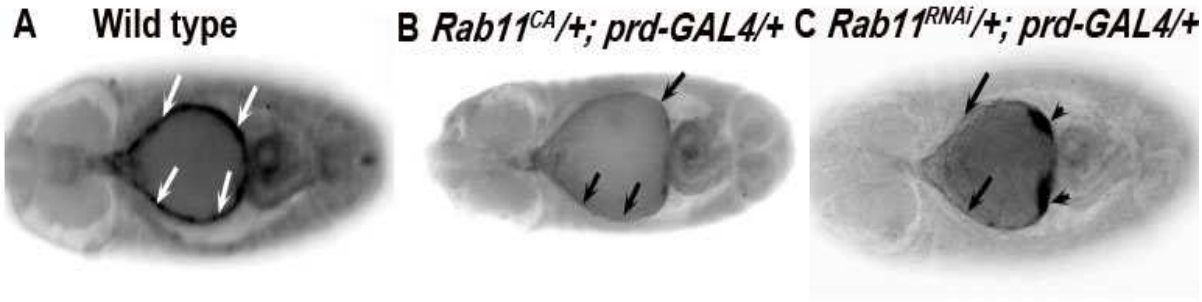

Fig 2. Immunostaining of stage 14 wild type (A), $Rab11^{CA}$ (B) and $Rab11^{RNAi}$ (C) embryos using anti-βPS-antibody. Anterior is on the left side in all the images. The arrows in B and C indicate the areas where there is a loss of βPS-integrins from the leading edge epidermal cells as compared with wild type. The arrowheads in C indicate the areas where there is an aggregation of βPS-integrins.

[Downloaded from ijmcmed.org on 2026-10-15]

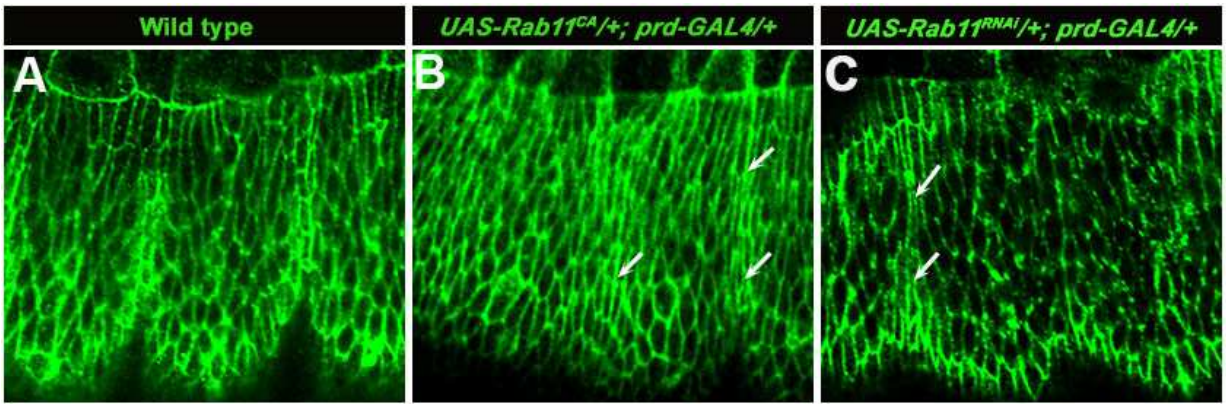

Fig 3. Confocal sections of wild type(A), *Rab11*^{CA}(B) and *Rab11*^{RNAi} (C) embryos, respectively, stained with anti-phosphotyrosine antibody showing the distribution of tyrosine phosphorylated proteins in the leading edge and the lateral epidermal cells. The dorsal side is up in all the images. The arrows in B and C indicate the change of cell shape in the lateral epidermal cells when compared to the wild type embryo (A).

Loss of Rab11 function results in change of lateral epithelial cell shape

It has been shown that integrin mediated cell-ECM adhesion is essential to maintain proper cell shape (12). We have recently shown that trafficking of βPS integrin through Rab11 endosomes is essential for cell adhesion, maintenance of cell shape and actin-cytoskeleton organisation during *Drosophila* wing development (14). To check the cell shape of the lateral epidermal cells in these two *Rab11* mutant backgrounds, immunostaining of the embryos was carried out using an apical marker, anti-phosphotyrosine antibody (19). This showed a change in the cell shape (Fig. 3B and 3C) when compared to the normal wild type embryos (Fig. 3A).

This study demonstrates that in *Rab11* mutant background there is a loss of βPS-integrin from the leading edge cells and there is change in the cell shape of the lateral epidermal cells. Thus this shows that Rab11 is essential for the maintenance of βPS-integrin level in the leading edge epidermal cells and the proper maintenance of cell shape in the lateral epidermal cells. Previous studies in *Drosophila* wing development have shown that cell-ECM interactions mediated by integrins regulate the change in cell shape which is essential for morphogenesis (12). Since defects are seen in localisation of βPS-integrin in the leading edge cells of *Rab11* mutants, through this study we indirectly demonstrate that Rab11 is essential for the trafficking of βPS-integrin and thereby proper maintenance of cell shape. This also supports the previous study done in *Drosophila* wing development (14).

Acknowledgements

We thank M. Gonzales-Gaitan, D. Ready, and the Bloomington Stock Centre for the fly stocks. The use of the national scanning confocal microscopy facility from DST is also gratefully acknowledged.

References

1. Lazar T, Gotte M, Gallwitz D. Vesicular transport: how many Ypt/Rab-GTPases make a eukaryotic cell? Trends Biochem Sci 1997;22:468-72.

2. Novick P, Zerial M. The diversity of Rab proteins in vesicle transport. Curr Opin Cell Biol 1997;9:496-504.

3. Pelissier A, Chauvin JP, Lecuit T. Trafficking through Rab11 endosomes is required for cellularization during *Drosophila* embryogenesis. Curr Biol 2003;13:1848-57.

4. Riggs B, Fasulo B, Royou A, et al. The concentration of Nuf, a Rab11 effector, at the microtubule-organizing center is cell cycle regulated, dynein-dependent, and coincides with furrow

[Downloaded from ijmcmed.org on 2026-10-15]

formation. Mol Biol Cell 2007;18:3313-22.

5. Bhuin T, Roy JK. Rab11 is required for myoblast fusion in *Drosophila*. Cell and Tissue Research 2009;336:489-99.

6. Bhuin T, Roy J. Rab11 is required for embryonic nervous system development in *Drosophila*. Cell Tissue Res 2009;335:349-56.

7. Campos-ortega JA, Hartenstein V. The embryonic development of *Drosophila melanogaster*. New York Springer:Verlag; 1985.

8. Noselli S .JNK signaling and morphogenesis in *Drosophila*. Trends Genet 1998;14:33-8.

9. Sasikumar S, Roy JK. Developmental Expression of Rab11, a Small GTP-Binding Protein in *Drosophila* Epithelia. Genesis 2009;47:32-9.

10. Hynes RO. Integrins: versatility, modulation, and signaling in cell adhesion. Cell 1992;69:11-25.

11. Hynes RO. Integrins: Bidirectional, allosteric signaling machines. Cell 2002;110:673-87.

12. Dominguez-Gimenez P, Brown NH, Martin-Bermudo MD. Integrin-ECM interactions regulate the changes in cell shape driving the morphogenesis of the *Drosophila* wing epithelium. J Cell Sci 2007;120:1061-71.

13. Narasimha M, Brown NH. Novel functions for integrins in epithelial morphogenesis. Curr Biol 2004;14:381-5.

14. Bhuin T, Roy JK. Rab11 is required for cell adhesion, maintenance of cell shape and actin-cytoskeleton organization during *Drosophila* wing development. Int J Dev Biol 2011;55:269-79.

15. Satoh AK, O'Tousa JE, Ozaki K, et al. Rab11 mediates post-Golgi trafficking of rhodopsin to the photosensitive apical membrane of *Drosophila* photoreceptors. Development 2005;132:1487-97.

16. Emery G, Hutterer A, Berdnik D, et al. Asymmetric Rab11 endosomes regulate delta recycling and specify cell fate in the *Drosophila* nervous system. Cell 2005;122:763-73.

17. Van VD, Kopczynski C. Techniques for anatomical analysis of the *Drosophila* embryonic nervous system. In: Richter J (ed). In Comparative methods to study the oocytes and embryos. New York: Oxford University Press Inc; 1999:490-513.

18. Brand AH, Perrimon N. Targeted Gene-Expression as a Means of Altering Cell Fates and Generating Dominant Phenotypes. Development 1993;118:401-15.

19. Muller HAJ, Wieschaus E. armadillo, bazooka, and stardust are critical for early stages in formation of the zonula adherens and maintenance of the polarized blastoderm epithelium in *Drosophila*. J Cell Biol 1996;134:149-63.

[Downloaded from ijmcmed.org on 2026-10-15]