Salvia officinalis (Sage) Leaf Extract as Add-on to Statin Therapy in Hypercholesterolemic Type 2 Diabetic Patients: a Randomized Clinical Trial

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The efficacy and safety of Salvia officinalis combined with statin have not been evaluated in dyslipidemic diabetes mellitus type 2 (DDMT2) so far. The plant extract antioxidant activity was determined by the DPPH radical scavenging assay. The total flavonoid, total phenolic and quercetin contents of the capsules containing the plant extract were also measured. Moreover, the effects of 2-month extract intake (500 mg capsule three times a day) as add-on to daily use of 15 mg glyburide, 2000 mg metformin and 10 mg atorvastatin on the blood levels of fasting glucose (FG), 2 h postprandial glucose (2hPPG), glycosylated hemoglobin (HbA1c), total cholesterol (TC), low-density lipoprotein cholesterol (LDL-C), triglyceride, high-density lipoprotein cholesterol (HDL-C), serum aspartate aminotransferase (AST), serum alanine aminotransferase (ALT), creatinine and body mass index were studied in 50 patients and compared with the placebo group (n=50). The extract IC₅₀ in the DPPH assay was 87.26±0.003 µg/mL (mean±SD), whereas the ascorbic acid IC₅₀ was 5.626±0.001 µg/mL (mean±SD). The total flavonoid, total phenolic and quercetin contents of the capsule containing the plant extract were 39.76±3.58 mg of rutin equivalents (mean±SD), 30.33±1.23 mg of gallic acid (mean±SD) and 0.13 mg, respectively. The extract lowered FG, 2hPPG, HbA1c, TC, LDL-C and triglyceride levels, but increased HDL-C level compared to the placebo at the endpoint (P<0.05). The extract did not affect the other parameters significantly and no adverse effect was reported. The extract has substantial antioxidant activity which may be beneficial for the prevention of the cardiovascular complications of DDMT2. Moreover, addition of the extract to statin therapy is apparently safe and further improves lipid profile.

Keywords: Salvia officinalis, statin, dyslipidemia, diabetes mellitus

Dyslipidemic diabetes mellitus type 2 (DDMT2) is common worldwide. It is characterized by blood lipid changes as raised total cholesterol, low-density lipoprotein cholesterol (LDL-C) and triglyceride and decreased level of high-density lipoprotein cholesterol (HDL-C) alongside hyperglycemia. Also, it is a major cause of atherosclerotic cardiovascular disease (CVD).
Co-administration of several antihyperglycemic drugs is usually necessary for the management of the hyperglycemia. Nevertheless, standard antihyperglycemic therapies do not have sufficient efficacy and safety. Many patients do not respond adequately to the therapies. Therefore, novel antihyperglycemic agents with more efficacy and safety are needed (1-4). LDL-C is the most important target in therapy of dyslipidemia. The use of statins is the preferred treatment strategy for both primary and secondary prevention of CVD. However, some patients respond inadequately, about 15% are intolerant, and other factors prevent achieving cholesterol goals in as many as 40% of patients. Thus, there is great need for additional drugs to prevent and treat CVD, whether as monotherapy or in combination with statin drugs (2-7). The plant kingdom can be a source of new anti-diabetic and anti-dyslipidemic drugs (8).

Plants with agonistic action on peroxisome proliferator-activated receptor γ (PPAR γ) may be useful in the treatment of dyslipidemia and diabetes mellitus type 2 (9). Agonists of PPAR γ prevent atherosclerotic CVD (10). Salvia officinalis L. (sage) (Lamiaceae family) leaves are used widely as a food flavoring. Moreover, infusion of 4-6 g of dry sage leaves in two divided doses per day is taken in the traditional medicine as an antihyperglycemic agent to treat diabetes mellitus (11). Sage consists of volatile oils, tannins, diterpenes, triterpenes, steroids, flavones and flavonoids (12-14). Sage leaf extracts (constituents) have demonstrated a variety of pharmacological effects such as in vitro agonistic action on the PPAR γ (12, 13), in vitro α-glucosidase and pancreatic lipase inhibition, and lipid absorption inhibitory and antihypertriglyceridemic effects in mice (14, 15), antioxidant and lipid peroxidation inhibitory effects in rat brain and liver homogenates in vitro (16), anti-inflammatory effect in rat hind paw acute inflammation induced with oil of turpentine (17), antihyperglycemic effect in streptozocin-induced diabetic rats (18), insulin sensitizer and gluconeogenesis inhibitory effects in primary cultures of hepatocytes from healthy sage-tea-drinking rats (metformin-like effects) (19). There have been few clinical trials on the antihyperglycemic and antidysslipidemic effects of sage (20-23). Notably, the efficacy and safety of sage combined with statin therapy to improve lipid profile have not been evaluated so far. Hence, the present study was undertaken. Since antioxidant property is clinically important for the prevention of the cardiovascular complications of DDMT2 (24), the radical scavenging activity of the extract was also studied. Moreover, the extract was standardized by determination of the total flavonoid, total phenolic, and quercetin contents.

Materials and methods

Preparation of the extract and placebo capsules

Sage was collected and extracted, and the extract and placebo capsules were prepared as described previously (22). The sage capsules contained 500 mg of the extract powder. The placebo capsules contained toast powder. The sage and placebo capsules were identical in appearance.

The extract DPPH assay

Inhibition of diphenyl-2-picrylhydrazyl (DPPH) radicals by the extract was assayed according to a method described previously (25). A mixture consisting of an extract solution at different concentrations (1.5 mL) and the methanolic solution of the DPPH reagent (1.5 mL) was prepared in a volumetric flask. The mixture was left to stand for 30 min in a dark place, and then the absorption was measured at 517 nm using a spectrophotometer (Human, USA). The radical scavenging activity (RSA) was calculated as a percentage of DPPH discoloration using the equation: RSA (%) = (Ac-As/Ac)×100 where Ac is the absorbance of the negative control and As is the absorbance of the plant sample or ascorbic acid.

Ascorbic acid was used as reference standard.
The assay results were expressed as IC50 denoting the antioxidant concentration which reduces the DPPH radicals about 50%.

**Determination of the extract total phenolic content**

The total phenolic contents were determined through the Folin-Ciocalteu colorimetric method described previously (26). In brief, the plant extract solution (1mL) was mixed with 500 µL of the Folin-Ciocalteu reagent and 5 mL distilled water in a volumetric flask. After 5 min, 1 mL of 15% sodium carbonate solution was added to the mixture and then kept in the dark for 30 min, after which the absorbance was determined at 725 nm using a spectrophotometer (Human, USA). Gallic acid was used to generate the standard curve, and the reduction of the Folin-Ciocalteu reagent by the samples was expressed as mg of gallic acid equivalents per capsule.

**Determination of the total flavonoid content**

The total flavonoid content was determined by a modified method described previously (27). The extract (1 mg/mL) or standard was mixed with 4 mL of distilled water and 300 µL of a 5% sodium nitrite solution and after 5 min, 300 µL of 10% aluminum chloride solution was added to the mixture. After 6 min, 2 mL of 1 M sodium hydroxide and 3 mL of distilled water were added to the mixture. The solution was properly mixed and absorbance was measured at 510 nm using a spectrophotometer (Human, USA). Rutin (100 µg/mL up to 1200 µg/mL) was used to construct the standard curve and the results were expressed as mg of rutin equivalents per capsule.

**Determination of the extract quercetin content**

Quercetin was measured by the HPLC method described previously (28). The chromatographic separations were achieved using a YMC triart C18 column (250 mmx4.6 mm, 5 µm). A reverse phase HPLC assay was carried out using an isocratic elution with a flow rate of 1 mL/min, a mobile phase of 35: 65 (acetonitrile: 0.2 phosphoric acid) and a detection wavelength of 360 nm. The injection volume was 50 µL of each solution. The total run time was 16 minutes for each injection. The solvents and distilled water were filtered previously through a PTF membrane using a set of glass bottles with the aid of a vacuum pump. The result was expressed as mg of quercetin per capsule.

**Protocol of the clinical trial**

A 2-arm, randomized, double-blind, placebo-controlled parallel-group trial with the following eligibility criteria was performed in the Diabetes Clinic (Karaj City, Alborz Province of Iran). Recruitment was carried out from April 20, 2012 to October 30, 2015. Inclusion criteria consisted of Iranian male and female type 2 diabetic patients aged 40-60 years with levels of HbA1C below 8% and LDL-C between 100 and 130 mg/dL despite daily intake of 15 mg glyburide, 2000 mg metformin and 10 mg atorvastatin. Patients with cardiovascular, pulmonary, renal and hepatic diseases; pregnant and breast feeding women were excluded from the study. 152 patients were screened. The enrolled patients were equally randomized to the extract and placebo groups. Random allocation was performed by block randomization using random number table. The patients were instructed to take one extract or placebo capsule every 8 h for 2 months. The patients’ treatment, diet and physical activity remained unchanged throughout the trial. The patients were requested to report any adverse drug reaction. At the beginning and endpoint of the trial, the patients’ blood levels of fasting glucose, 2 h postprandial glucose, HbA1c (glycosylated hemoglobin), total cholesterol, triglyceride, LDL-C, HDL-C, serum aspartate aminotransferase (AST), serum alanine aminotransferase (ALT) and creatinine were determined using standard enzymatic kits produced by Pars Azmoon company (Tehran, Iran) and an auto-analyzer (Hitachi 902, Japan).
The patients’ body mass index (BMI) was also measured at the baseline and endpoint. Fasting glucose, HbA1c and LDL-C were the primary outcome measures. The other blood variables and BMI were the secondary outcome measures. Compliance was measured by counting returned drugs and asking how many doses of the drugs were (or were not) taken. 50 patients in each group was the sample size calculated to detect 20 mg/dL difference of fasting glucose level between the groups, considering type I error= 0.05 and 80% power. The chi-squared and independent samples t tests were used for data analysis and p<0.05 was considered as significant. SPSS 20 was used for statistical analysis. The data were analyzed by the intention-to-treat approach. The Ethics Committee of the Endocrinology and Metabolism Research Institute affiliated to the Tehran University of Medical Sciences approved the protocol. The trial was performed in accordance with the revised declaration of Helsinki 2013. The patients signed a written informed consent form before enrollment. The trial was registered in the Iranian Registry of Clinical Trials with the number IRCT201506082288N7.

**Results**

**Phytochemical analysis of the extract**

The IC\textsubscript{50} of the extract was 87.26± 0.003 µg/mL (mean±SD), whereas the IC\textsubscript{50} of ascorbic acid was 5.626 ± 0.001 µg/mL (mean ± SD). The total flavonoid content as milligram rutin equivalent per capsule was 39.76± 3.58 (mean± SD). The total phenolic content of the extract as milligrams of gallic acid per capsule was 30.53± 1.23 (mean± SD). The aforementioned values are means of 3 measurements. Moreover, the amount of quercetin in each capsule containing the plant extract was 0.13 mg. The figures 1 and 2 demonstrate the HPLC chromatograms of the test sample and standard quercetin.

**Clinical trial**

![Fig. 1. HPLC chromatogram of the test sample.](image1)

![Fig. 2. HPLC chromatogram of the standard quercetin.](image2)
105 patients entered the study. 50 patients in each of the sage and placebo groups completed the trial. The CONSORT flow diagram of the trial is shown in the figure 3. The patients fully complied with the protocol. Demographic data of the patients in the sage and placebo groups did not differ significantly (Table 1). There was no significant difference between the sage and placebo groups in terms of the blood parameters levels and BMI at the baseline. The extract significantly lowered the fasting glucose, 2 h postprandial glucose, HbA1c, total cholesterol, triglyceride and LDL-C and increased HDL-C in DDMT2 patients. Specifically, the current study demonstrated the benefit of sage added to the background statin therapy. Sage as add-on to statin therapy was apparently safe and further improved lipid profile in DDMT2 patients. The sage effects on the glycemic control and lipid profile may ^

Discussion

The main findings of the present clinical trial were that sage combined with glyburide, metformin and atorvastatin further lowered fasting and 2 h postprandial glucose, HbA1c, total cholesterol, triglyceride and LDL-C and increased HDL-C in DDMT2 patients. Specifically, the current study demonstrated the benefit of sage added to the background statin therapy. Sage as add-on to statin therapy was apparently safe and further improved lipid profile in DDMT2 patients. The sage effects on the glycemic control and lipid profile may ^
Table 1. Demographic characteristics of the study participants

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Sage group</th>
<th>Placebo group</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gender</td>
<td>21 males, 29 females</td>
<td>22 males, 28 females</td>
</tr>
<tr>
<td>Age (years)</td>
<td>54.77± 4.82</td>
<td>52.57± 5.69</td>
</tr>
<tr>
<td>Duration of diabetes mellitus type 2 (years)</td>
<td>10.6± 3.6</td>
<td>9.8± 4.2</td>
</tr>
</tbody>
</table>

Where appropriate, values are given as mean± standard deviation.

Table 2. The blood parameter levels and body mass index before and after intervention

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Mean (SD) before intervention</th>
<th>P-value</th>
<th>Mean (SD) after intervention</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fasting glucose (mg/dL)</td>
<td>177.8 (32.4) S</td>
<td>0.171</td>
<td>158.3 (26.6) S</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td></td>
<td>188.8 (38.9) P</td>
<td></td>
<td>197.5 (46.4) P</td>
<td></td>
</tr>
<tr>
<td>2 hours postprandial glucose (mg/dL)</td>
<td>207.0 (43.9) S</td>
<td>0.609</td>
<td>158.3 (26.6) S</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td></td>
<td>202.0 (43.2) P</td>
<td></td>
<td>206.8 (43.9) P</td>
<td></td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>7.2 (1.1) S</td>
<td>0.320</td>
<td>6.7 (0.9) S</td>
<td>0.007</td>
</tr>
<tr>
<td></td>
<td>7.0 (1.1) P</td>
<td></td>
<td>7.3 (1.1) P</td>
<td></td>
</tr>
<tr>
<td>Total cholesterol (mg/dL)</td>
<td>228.2 (42.9) S</td>
<td>0.273</td>
<td>182.6 (36.8) S</td>
<td>0.042</td>
</tr>
<tr>
<td></td>
<td>216.6 (50.4) P</td>
<td></td>
<td>203.6 (52.9) P</td>
<td></td>
</tr>
<tr>
<td>Triglyceride (mg/dL)</td>
<td>274.1 (83.5) S</td>
<td>0.176</td>
<td>179.1 (52.8) S</td>
<td>0.020</td>
</tr>
<tr>
<td></td>
<td>245 (105.7) P</td>
<td></td>
<td>220 (98.1) P</td>
<td></td>
</tr>
<tr>
<td>LDL-C (mg/dL)</td>
<td>117.2 (9.1) S</td>
<td>0.776</td>
<td>109.3 (29.7) S</td>
<td>0.012</td>
</tr>
<tr>
<td></td>
<td>116.7 (8.1) P</td>
<td></td>
<td>123.5 (18.4) P</td>
<td></td>
</tr>
<tr>
<td>HDL-C (mg/dL)</td>
<td>45.4 (11.1) S</td>
<td>0.823</td>
<td>50.5 (9.6) S</td>
<td>0.028</td>
</tr>
<tr>
<td></td>
<td>44.8 (9.7) P</td>
<td></td>
<td>45.7 (9.3) S</td>
<td></td>
</tr>
<tr>
<td>AST (U/L)</td>
<td>21.2 (6.2) S</td>
<td>0.699</td>
<td>22.7 (13) S</td>
<td>0.714</td>
</tr>
<tr>
<td></td>
<td>21.7 (5.9) P</td>
<td></td>
<td>23.8 (13.2) P</td>
<td></td>
</tr>
<tr>
<td>ALT (U/L)</td>
<td>23.6 (9.2) S</td>
<td>0.534</td>
<td>24.9 (6.4) S</td>
<td>0.717</td>
</tr>
<tr>
<td></td>
<td>24.8 (7.5) P</td>
<td></td>
<td>25.5 (8.2) P</td>
<td></td>
</tr>
<tr>
<td>Creatinine (mg/dL)</td>
<td>0.9 (0.1) S</td>
<td>0.169</td>
<td>1 (0.2) S</td>
<td>0.209</td>
</tr>
<tr>
<td></td>
<td>0.9 (0.1) P</td>
<td></td>
<td>0.9 (0.1) P</td>
<td></td>
</tr>
<tr>
<td>Body mass index (kg/m²)</td>
<td>27.7 (4.9) S</td>
<td>0.782</td>
<td>26.6 (7) S</td>
<td>0.618</td>
</tr>
<tr>
<td></td>
<td>28.1 (6.5) P</td>
<td></td>
<td>27.5 (6) P</td>
<td></td>
</tr>
</tbody>
</table>

P< 0.05 is significant (independent samples t-test). S: sage group; P: placebo group; SD: standard deviation.

Flavonoids, total phenolics and quercetin. These compounds may be responsible for the antioxidant effect of sage. The sage effects concur with the previous clinical trials (20-23). In a crossover trial with 6 healthy female volunteers (aged 40-50), 4-week sage tea drinking (infusion prepared by pouring 300 ml of boiling water onto 4 g dried leaves of sage, twice a day) lowered blood LDL-C, total cholesterol and LDL-C/HDL-C but increased blood HDL-C without hepatotoxic or other adverse effects. However, the effects of sage on the blood triglyceride and VLDL levels were not evaluated in.
the trial. Moreover, the subjects were normo-
lipidemic, the trial had a small sample size, and was
not randomized, double-blind and placebo-
controlled (20). In a 2-month randomized double-
blind placebo-controlled trial, sage leaf (80%) ethanolic extract (500 mg t.i.d.) lowered blood total cholesterol, triglyceride and VLDL levels, but increased blood HDL-C in 40 hyperlipidemic (hypercholesterolemic and/or hyper-
triglyceridemic) patients not receiving any other
antihyperlipidemic agent (21). In another 3-month
randomized double-blind placebo-controlled trial, sage leaf 80% ethanolic extract (500 mg t.i.d.) reduced blood fasting glucose, HbA1c, total
cholesterol, LDL-C and triglyceride, but increased
blood HDL-C in 40 hyperlipidemic (hypercholes-
teroemic and/or hypertriglyceridemic) type 2 diabetic patients taking 10 mg glyburide and 1000 mg metformin daily, not receiving any other
antihyperlipidemic agent (22). Besides, according
to a randomized double-blind trials, intake of 150
mg sage extract tablet t.i.d. combined with standard antidiabetic treatment by 40 type 2 diabetic patients reduced 2 h postprandial glucose and total
cholesterol levels without affecting fasting glucose,
HbA1c and other lipid parameters compared to the
placebo group (n= 40) after 3 months (23). Inadequate sage extract dose and small sample size are the main limitations of the just mentioned study. Comparison of the inclusion criteria and baseline
glycemia and lipid levels of the present trial with
those of the previous trials (21-23) indicates that the
participants of this study had more severe hyperglycemia and hyperlipidemia (hypercholesterol- emia and hypertriglyceridemia). The present
study validates the traditional use of sage for the
treatment of diabetes mellitus. There is little data on
the constituents and mechanisms mediating the
antihyperglycemic and antidysholipidemic effects of
sage. Liposoluble compounds including the phenols
carnosic acid and carnosol together with unknown
hydrosoluble ingredients as well as PPARγ
agonistic and metformin-like actions and α-
glucosidase, pancreatic lipase and lipid absorption
inhibition have been implicated in the sage effects
(12, 13, 15, 19, 21, 22).

Finally, sage has promising pharmacological
effects which justify more research. Conduction of
more clinical trials regarding the effects of sage in
the treatment of dyslipidemias and diabetes mellitus
as well as identification of the compounds and
mechanisms involved in the sage effects are
warranted.

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Conflicts of Interest
The authors declared no conflict of interest.

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