IJMCM
Autumn 2015, Vol 4, No 4

Original Article

Association Between *Helicobacter pylori* cagA, babA2 Virulence Factors and Gastric Mucosal Interleukin-33 mRNA Expression and Clinical Outcomes in Dyspeptic Patients

Heshmat Shahi[1], Somayeh Reiisi[2], Rasol Bahreini[3], Nader Bagheri[1], Loghman Salimzadeh[1]
Hedayatollah Shirzad[1*]

1. Cellular and Molecular Research Center, Shahrekord University of Medical Sciences, Shahrekord, Iran.
2. Department of Genetic, Faculty of basic science, Shahrekoed University, Shahrekord, Iran.
3. Department of Internal Medicine, Shahrekord University of Medical Sciences, Shahrekord, Iran.

Submmited 25 November 2015; Accepted 16 December 2015; Published 23 December 2015

Helicobacter pylori (*H. pylori*) infection has been reported in more than half of the world human population. It is associated with gastric inflammation and noticeable infiltration of the immune cells to the stomach mucosa by several cytokines secretion. IL-1β, IL-18 have been shown to contribute to *H. pylori* induced gastritis, but the details of inflammation and association of virulence factors remain unclear. IL-1 cytokine family has a new additional cytokine, Interleukin-33 (IL-33), which is contemplated to have an important role for host defense against microorganisms. *H. pylori* virulence factors important in gastritis risk are the cag pathogenicity island (cag-PAI) and babA. This study evaluated IL-33 mucosal mRNA expression levels in infected and uninfected patients and its relationship with bacterial virulence factors cagA, babA$_2$ and type of gastritis. Total RNA was extracted from gastric biopsies of 79 *H. pylori*-infected patients and 51 *H. pylori*-negative patients. Mucosal IL-33 mRNA expression levels in gastric biopsies were assessed using real-time PCR. Existence of virulence factors were detected by PCR. IL-33 mRNA expression was significantly higher in biopsies of *H. pylori*-infected patients compared to *H. pylori*-uninfected patients (P<0.0001). Also there was a direct relationship between virulence factor bab-A$_2$ and enhancement in IL-33 mRNA expression. Furthermore, IL-33 mRNA expression level was significantly lower in chronic gastritis patients compared with patients with active gastritis (P<0.001). IL-33 may play a crucial role in the inflammatory response and induction of the chronic gastritis and severity of inflammatory changes in the gastric mucosa.

Key words: *Helicobacter pylori*, gastritis, interleukin-33, virulence factor

Helicobacter pylori (*H. pylori*) is a Gram-negative bacterium that attaches to the stomach epithelial cells and colonizes in the stomach of approximately half of the world population, causing long lived gastric inflammation despite the host immune responses. (1, 2). *H. pylori*

* Corresponding author: . Cellular and Molecular Research Center, Shahrekord University of Medical Sciences, Shahrekord, Iran.
E-mail: Shirzadeh@yahoo.com

[Downloaded from ijmcmed.org on 2026-09-16]

infection is associated with gastritis and immune cell infiltration into the gastric mucosa that may lead to chronic gastritis, peptic ulcer and production of pro inflammatory cytokines such as IL-1, IL-8, and TNFα (3, 4). In recent years, a newly addition to the IL-1 family is IL-33 which has high similarity to IL-18. In contrast to IL-18, IL-33 has anti-inflammatory effects although a TH1 response has been reported (5, 6). IL-33 is produced as a 30 kD protein which is cleaved by caspase to produce an 18 kD form. IL-33 is highly expressed in the stomach epithelium. However, its gastric function is unknown (5). IL-33 exists in the nucleus and cytoplasm of macrophages, dendritic cells, fibroblast and endothelial cells (7). The role of IL-33 and its receptors ST2 "IL-33/ST2" have been identified as crucial to the homeostasis of the epithelial inflammation specially in intestinal epithelium (8). Patients with ulcerative colitis show increased IL-33 levels in the epithelium of intestine (9). The ST2 gene relates to the IL-1/TLRs super family which produces four ST2 protein isoforms (10). In addition, serum ST2 levels are associated with severity of disease that may be a biomarker of disease degree (7, 11). Some researchers showed high expression level of IL-33 in endothelial cells and cancer cell lines (8, 12). IL-33 is known to be associated with inflammatory tissue in crohn's disease and rheumatoid arthritis (9). Although virulence factors may induce gastric inflammation, atrophy, metaplasia and malignancy in the stomach, it has been reported that mucosal levels of pro-inflammatory cytokines in the infection site with *H. pylori* are correlated with the various *H. pylori* virulence factors (13). This study evaluated IL-33 mucosal mRNA expression levels in infected and uninfected patients and assessed its relationship with bacterial virulence factors cagA, babA$_2$ and type of gastritis.

Material & methods

Patients & sampling

130 specimens were collected from patients presenting dyspepsia symptoms and gastrointestinal disorders. The process was approved by Ethics Committee of Shahrekord University of Medical Sciences. Gastric biopsy specimens were taken from the antrum (pyloric gland area). Gastritis was investigated by endoscopy. None of the patients had received anticoagulants and nonsteroidal anti-inflammatory drugs (NSAIDs) for 1 month before specimen collection and none of them had received treatment for *H. pylori* infection and no autoimmune disease was reported (14). 79 *H. pylori* infected gastritis patients including 40 men (40.02 ± 15.65 years) and 39 women (38.9 ± 13 years) and 51 uninfected gastritis patients, 23 men (41.42 ± 12.25 years) and 28 women (39.8 ± 15.02 years), contributed to this study. *H. pylori* infection was detected by the rapid urease test (RUT), polymerase chain reaction (PCR), and pathological examination (PE) of three biopsies taken from the antrum. Patients with all positive tests (RUT, PCR, PE) were considered as positive for *H. pylori* infection. Determination of bacterial virulence factors was conducted by PCR test and one biopsy from each case was used for measuring IL-33 mRNA expression rate by real-time PCR.

Histological examination

Gastric biopsy specimens were merged in 10% buffered formalin and stained with hematoxylin and eosin (H&E) to grade gastritis and with giemsa for *H. pylori* detection. The severity of gastritis was graded from 1-3 (Mild, Moderate, Severe) based on the degree of immune cells infiltration, polymorphonuclear leukocyte (PMN) and mononuclear cell (MNC) infiltration, and dysmorphic according to the updated Sydney system (15).

Molecular characterization of *H. pylori*

Genomic DNA from all samples was extracted by Biospin tissue genomic DNA extraction kit (BioFlux, Japan) PCR. Specific primers for *H. pylori* and its virulence factors PCR test were: 16S rRNA, forward: 5'-CTGGAGAGACTAAGCCCTC

[Downloaded from ijmcmed.org on 2026-09-16]

C-3' and reverse: 5'-ATTACTGACGCTGATTGTG C-3', glmM (ureA), forward: 5'-AAGCTTTTAG GGGTGTTAGGGGTTT-3' and reverse: 5'-AAGC TTACTTTCTAACACTAACGC-3' (housekeeping and specific genes, respectively), cagA, forward: 5'-ATGACTAACGAAACTATTGATC-3' and reverse :5'-CAGGATTTTTGATCGCTTTATT-3', babA2, forward:5'- CCAAACGAAACAAAAAGCGT - 3' and reverse: 5'-GCTTGTGTAAA AGCCGTCGT-3' . For cagA, babA$_2$ gene evaluation, the PCR program contained 35 cycles of denaturation (94 °C for 30 s), annealing (56 °C for 30 s, extension at 72 °C for 30 s), and one final extension (72 °C for 5 min) (16, 17).

Analysis of IL-33 mRNA expression in the gastric mucosa samples by real-time PCR

Total RNA was extracted from gastric biopsy samples by total RNA extraction biozol (bioflux, Japan). An aliquot containing 0.1 mg of total RNA was used for the reverse transcription (RT) reaction, according to the manufacturer's instructions first-strand cDNA synthesis system (Takara, Japan). The sequences of oligonucleotide primers and probe designed by Oligo.7 software for β-actin and IL-33 are: β-actin, forward 5'-AGCCTCGCCTTTGC-CGA-3' and reverse 5' -CTGGTGCCTGGGGCG-3' and probe FAM 5'-CCGCCGCCCGTCCACACC-CGCC-3' TAMRA, for IL-33, forward 5'-TGGAGGATGAAAGTTATGAG-3', reverse 5'-TCAGGGTTACCATTAACATC-3', probe FAM 5'-TACCATCAACACCGTCACCTGATTCA-3' TAMRA. The assessment of IL-33 mRNA levels was performed using a Rotor-Gene 3000 (Corbett, Australia). real-time PCR reactions were done in a total volume of 25 µl containing 12.5 µl of 2x Rotor-Gene Probe PCR Master Mix (Qiagen, Germany), 3 µl of synthesized cDNA solution, 500 nM of each forward & reverse primer and 250 nM of the TaqMan probe. Amplification process included a primary warming step (94 °C for 10 min), denaturation step (94 °C for 15 s) and an annealing/extension step (60 °C for 60 s). Relative quantification of cytokine mRNA to β-actin mRNA was determined using the $2^{-\Delta}$Ct method (18). Each assay was done in duplicate.

Statistical analyzes

In statistics, normality tests were used to determine whether a data set is well-modeled by a normal distribution or not. Cytokine expression is presented as means and differences between infected and uninfected groups were analyzed using the Mann-Whitney test. P values<0.05 were considered as significant.

Results

Genotype analysis

Table 1 gives an overview of the frequency distribution of the cagA, babA$_2$ status. The babA$_2$ gene was found in 49.5% of the *H. pylori* positive biopsies (Table 1). The cagA gene was detected in 68.3% (69 biopsies) of *H. pylori* positive specimens. The frequency of virulence factors in *H. pylori* positive biopsies is shown in table 2.

Table 1. Frequency of the cagA and the babA$_2$ in studied patients

Genotype	Frequency N	%
cagA		
Positive	69	68.3
Negative	32	31.7
babA$_2$		
positive	50	49.5
negative	51	50.5

[Downloaded from ijmcmed.org on 2026-09-16]

Table 2. Frequency of virulence factors in *H. pylori* positive biopsies

Genotype		Frequency	
cagA	$babA_2$	N	(%)
+	+	43	62.3
+	-	26	37.7
-	+	7	21.9
-	-	25	78.1

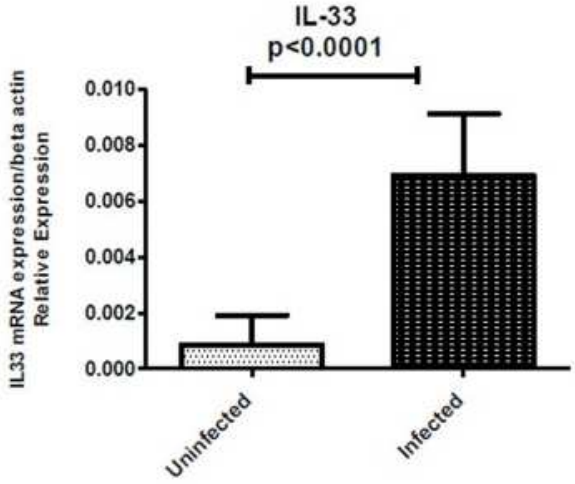

Fig. 1 . Mucosal IL-33 mRNA expression level in gastritis patients. 79 *H. pylori* infected and 51 *H. pylori* non-infected patients with gastritis were analyzed for IL-33 expression by real-time *PCR*. Levels are normalized to β-actin.

Elevated IL-33 expression in *H. pylori*-infected gastric mucosa

IL-33 mRNA level was detectable in all samples regardless of whether biopsies were taken from *H. pylori*-infected or uninfected patients. IL-33 expression was significantly higher in biopsy specimens of *H. pylori*-infected patients compared with uninfected patients (P<0.0001) (Fig. 1). If the mean *H. pylori*-positive group is 0.0069 and the mean *H. pylori*-negative group is 0.0008 then the difference in the IL-33 mRNA expression in the *H. pylori*-positive compared with the *H. pylori*-negative patients is 0.0069/0.0008 or 8.6 fold.

Effect of cagA and $babA_2$ on the mucosal IL-33 mRNA levels in *H. pylori*-infected patients

A mucosal IL-33 mRNA level was dependent on virulence factors status. If the mean level of IL-33 in *H. pylori*-cagA positive group is 0.00798 and the mean level of IL-33 in *H. pylori*-cagA negative group is 0.0079, then the difference in the IL-33 mRNA expression in the cagA-positive *H. Pylori* strains in comparison with the cagA-negative *H. pylori* strains biopsies is 0.00798/0.0079 or 1.01 fold. The difference in the IL-33 mRNA expression in the babA2-positive *H. pylori* strains compared with the $babA_2$-negative *H. pylori* strains is 0.0067/0.0012 or 5.58 fold (Fig. 2).

Association between mucosal IL-33 mRNA levels and gastric inflammation classification

During this study, semi-quantitative methods of scoring according to the updated Sydney system were used. The degree of active inflammation was evaluated and scored as below: 19.8% mild, 24.8% moderate and 19.8% severe, and more than 64% of gastritis patients were in an active stage of *H. pylori* gastritis. In addition the degree of chronic inflammation was assessed and graded as follow: 11.9% mild, 13.9% moderate, 9.9% severe, and

[Downloaded from ijmcmed.org on 2026-09-16]

more than 35% of gastritis patients were in a chronic phase of *H. pylori* gastritis (Table 3).

Association between mucosal IL-33 mRNA levels and type of gastritis

Figure 3 shows the relationship between the mucosal IL-33 mRNA expression and chronic inflammation (mononuclear cell infiltration) scores. There was a significant correlation between the mucosal IL-33 mRNA expression and the active inflammation scores (P<0.001). If the mean of IL-33 mRNA level in chronic gastritis group is 0.0031 and the mean of IL-33 mRNA level in active gastritis group is 0.0083 then the difference in the IL-33 mRNA expression in the active gastritis compared with the chronic gastritis biopsies is 0.0083/0.0031 or 2.67 fold (Fig. 3).

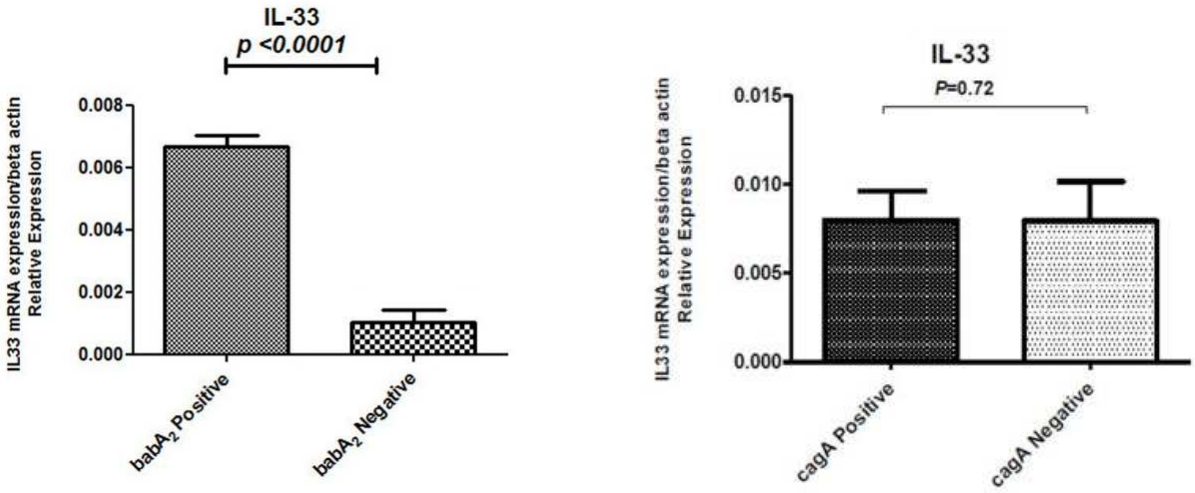

Fig. 2. Mucosal IL-33 mRNA expression in *H. pylori* infected patients according to virulence factors. IL-33 mRNA expression level was significantly lower in $babA_2$ negative gastritis patients compared with $babA_2$ positive patients. There is no significant relationship between IL-33 gene expression and existence of cagA virulence factor.

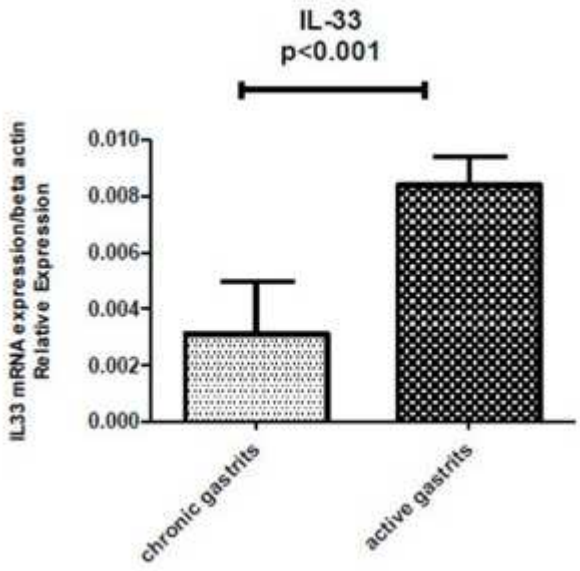

Fig. 3. Mucosal IL-33 mRNA levels according to gastritis status. IL-33 mRNA expression level was significantly lower in chronic gastritis patients compared with patients with active gastritis.

[Downloaded from ijmcmed.org on 2026-09-16]

Table. 3. *H.pylori* gastritis status according to updated Sydney classification

Description	Mild N (%)	Moderate N (%)	Severe N (%)	Total N (%)
Active	14 (19.8)	18 (24.8)	14 (19.8)	46 (64.4)
Chronic (no activity)	8 (11.6)	10 (14)	7 (10)	25 (35.6)
H.pylori positive	all	all	all	all
Man/woman	10/12	14/14	9/12	33/38
Total	22 (30.99)	28 (39.44)	21 (29.57)	71 (100)

Discussion

H. pylori infection may remain in the host for a long time and *H. pylori* pathogenesis is related to its virulence factors including cagA and babA. The cagA positivity in Iranian isolates varies between 44% to 91% in different reports. In the present study, 68.3% of the patients were infected with cagA- positive strains, similar to other Iranian reports (1, 19, 20). The babA positivity in Iranian isolates has been reported to vary from 40.6% to 94% in different studies. In the present study, 49.5% of the patients were infected with babA-positive strains, similar to other Iranian reports (1, 21, 22).The IL-1 family of cytokines has an established role in immune regulation and inflammatory processes. IL-33 has been observed to supply protection during gastric infection and may be an important mediator of the immune functions after damage or infection in epithelial cells (23, 24). IL-33 has been reported to reduce colonization and pathological effects in gastrointestinal infection (25). Recently several researches suggested that IL-33 may be involved in tumorogenesis and development of vascular diseases (26). In the present study, IL-33 mRNA expression was significantly higher in *H. pylori* infected patients than non-infected patients. Similar to our results, in another study in 2015, IL-33 mRNA expression increased with bacterial infection in stomach (27). However, in another study in 2015, IL-33 mRNA expression was reduced in chronic infection by *H. pylori* (14). In the present study, IL-33 mRNA expression was significantly high in $babA_2$ positive *H. Pylori* infected patients as well as active gastritis patients. To the best of our knowledge, this report is the first study considering the association between IL-33 mRNA expression and virulence factors. Like our results, another study in 2015 showed that IL-33 mRNA expression was increased in active gastritis biopsies more than chronic gastritis samples (14). The expression levels of other cytokines from IL-1 family such as IL-18 was higher in *H. pylori*-positive patients and cagA+ infected patients compared with *H. pylori*-negative patients and cagA negative infected patients(28). The results of a study in 2013, showed that there was no association between virulence factors and IL-18 mRNA expression (29). Another study showed that serum levels of IL-33 were significantly higher in patients with gastric cancer than healthy people, suggesting that serum IL-33 levels may have a closer correlation with gastric cancer than IL-18 levels. In other words, elevated serum IL-33 level was found to be an independent prognostic indicator (30). Stimulation through both Toll-Like receptors (TLRs) and Nod-Like receptors (NLRs) are all required for processing and release of IL-18 and IL-33 from normal monocytes (31). Similarly, in another study, elevated IL-33 mRNA expression was detected only in lipopolysaccharide (LPS)-activated monocytes (5). LPSs from Escherichia

[Downloaded from ijmcmed.org on 2026-09-16]

coli stimulate host immune cells via TLR4 and TLR2 and enhance IL-33 gene expression level in monocytes (32-34). Some data suggest that engagement of both TLR2 and TLR4 pathways stimulates IL-33 indicating a potential role for this cytokine in immune responses to diverse pathogens and pathogen associated microbial patterns (PAMPs) (35).

Our data suggest that reduction in IL-33 mRNA expression level may be a biomarker for predicting the prognosis of gastritis, especially in chronic gastritis which is an initiation for gastric ulcer. Prospective studies in a larger population and more virulence factors should be carried out to confirm the findings.

Conflict of interest

The authors declared no conflict of interests.

References

1. Shahi H, Reiisi S, Sadeghiani M, et al. Prevalence of cagA and babA2 genes in Helicobacter pylori strains Isolated from Iranian gastrointestinal disorder patients and their gastritis classification. Jbtw 2014;3:256-60.

2. Shahi H, Moghni M, Bahreini B, et al. Association between H.pylori babA virulence factor with clinical outcome and ABO blood group. JPAM 2015;9:285-90.

3. Labigne A, de Reuse H. Determinants of Helicobacter pylori pathogenicity. Infectious agents and disease 1996;5:191-202.

4. Dunn BE, Cohen H, Blaser MJ. Helicobacter pylori. Clinical microbiology reviews 1997;10:720-41.

5. J. S, Owyang A, Oldham E, et al. IL-33, an interleukin-1-like cytokine that signals via the IL-1 receptor-related protein ST2 and induces T helper type 2-associated cytokines. Immunity 2005;23:479-90.

6. Smithgall MD, Comeau MR, Yoon BR, et al. IL-33 amplifies both Th1- and Th2-type responses through its activity on human basophils, allergen-reactive Th2 cells, iNKT and NK cells. International immunology 2008;20:1019-30.

7. Nunes T, Bernardazzi C, de Souza HS. Interleukin-33 and Inflammatory Bowel Diseases: Lessons from Human Studies. Mediators Inflamm 2014;20:10-20.

8. Moussion C, Ortega N, Girard JP. The IL-1-like cytokine IL-33 is constitutively expressed in the nucleus of endothelial cells and epithelial cells in vivo: a novel 'alarmin'? PloS one 2008;3:e3331.

9. Carriere V, Roussel L, Ortega N, et al. IL-33, the IL-1-like cytokine ligand for ST2 receptor, is a chromatin-associated nuclear factor in vivo. Proceedings of the National Academy of Sciences of the United States of America 2007;104:282-7.

10. Bergers G, Reikerstorfer A, Braselmann S, et al. Alternative promoter usage of the Fos-responsive gene Fit-1 generates mRNA isoforms coding for either secreted or membrane-bound proteins related to the IL-1 receptor. The EMBO journal 1994;13:1176-88.

11. Garcia-Miguel M, Gonzalez MJ, Quera R, et al. Innate Immunity Modulation by the IL-33/ST2 System in Intestinal Mucosa. Biomed Res Int 2013;20:13-23.

12. Masamune A, Watanabe T, Kikuta K, et al. Nuclear expression of interleukin-33 in pancreatic stellate cells. American journal of physiology Gastrointestinal and liver physiology 2010;299:G821-32.

13. Kudo T, Nurgalieva ZZ, Conner ME, et al. Correlation between Helicobacter pylori OipA protein expression and oipA gene switch status. Journal of clinical microbiology 2004;42:2279-81.

14. Buzzelli JN, Chalinor HV, Pavlic DI, et al. IL33 Is a Stomach Alarmin That Initiates a Skewed Th2 Response to Injury and Infection. CMGH 2015;1:203-21.

15. Manxhuka-Kerliu S, Telaku S, Devolli-Disha E, et al. Helicobacter pylori gastritis updated Sydney classification applied in our material. Prilozi / Makedonska akademija na naukite i umetnostite, Oddelenie za bioloski i medicinski nauki = Contributions / Macedonian Academy of Sciences and Arts, Section of Biological and Medical Sciences 2009;30:45-60.

16. Lobo Gatti L, Agostinho Jn F, De Labio R, et al. Helicobacter pylori and cagA and vacA gene status in children from Brazil with chronic gastritis. Clin Exp Med 2003;3:166-72.

17. Sheu BS, Sheu SM, Yang HB, et al. Host gastric Lewis expression determines the bacterial density of Helicobacter pylori in babA2 genopositive infection. Gut 2003;52:927-32.

18. Schmittgen TD, Livak KJ. Analyzing real-time PCR data by the comparative C(T) method. Nature protocols 2008;3:1101-8.

[Downloaded from ijmcmed.org on 2026-09-16]

19. Jafari F, Shokrzadeh L, Dabiri H, et al. vacA genotypes of Helicobacter pylori in relation to cagA status and clinical outcomes in Iranian populations. Japanese journal of infectious diseases 2008;61:290-3.

20. Talebkhan Y, Mohammadi M, Mohagheghi MA, et al. CagA gene and protein status among Iranian Helicobacter pylori strains. Digestive diseases and sciences 2008;53:925-32.

21. Talebi Bezmin Abadi A, Taghvaei T, Mohabbati Mobarez A, et al. High correlation of babA 2 -positive strains of Helicobacter pylori with the presence of gastric cancer. Intern Emerg Med 2013;8:497-501.

22. Shirazi MH, Pakbaz Z, Douraghi M, et al. Frequency of babA2 genotype in Helicobacter pylori from Patient with Gastroduodenal Diseases in firouzgar hospital tehran. Govaresh 2012;17.

23. Duan L, Chen J, Zhang H, et al. Interleukin-33 ameliorates experimental colitis through promoting Th2/Foxp3(+) regulatory T-cell responses in mice. Molecular medicine 2012;18:753-61.

24. Grobeta P, Doser K, Falk W, et al. IL-33 attenuates development and perpetuation of chronic intestinal inflammation. Inflammatory bowel diseases 2012;18:1900-9.

25. Wills-Karp M, Rani R, Dienger K, et al. Trefoil factor 2 rapidly induces interleukin 33 to promote type 2 immunity during allergic asthma and hookworm infection. The Journal of experimental medicine 2012;209:607-22.

26. Choi YS, Choi HJ, Min JK, et al. Interleukin-33 induces angiogenesis and vascular permeability through ST2/TRAF6-mediated endothelial nitric oxide production. Blood 2009;114:3117-26.

27. Noto JM, Peek Jr RM. Sound the Alarmin: Interleukin-33 Commandeers the Gastric Immune Response. CMGH 2015;1:127-8.

28. Shimada M, Ando T, Peek RM, et al. Helicobacter pylori infection upregulates interleukin-18 production from gastric epithelial cells. European journal of gastroenterology & hepatology 2008;20:1144-50.

29. Bagheri N, Taghikhani A, Rahimian G, et al. Association between virulence factors of helicobacter pylori and gastric mucosal interleukin-18 mRNA expression in dyspeptic patients. Microbial pathogenesis 2013;65:7-13.

30. Sun P, Ben Q, Tu S, et al. Serum interleukin-33 levels in patients with gastric cancer. Dig Dis Sci 2011;56:3596-601.

31. Becker CE, O'Neill LA. Inflammasomes in inflammatory disorders: the role of TLRs and their interactions with NLRs. Seminars in immunopathology 2007;29:239-48.

32. Barksby HE, Nile CJ, Jaedicke KM, et al. Differential expression of immunoregulatory genes in monocytes in response to Porphyromonas gingivalis and Escherichia coli lipopolysaccharide. Clinical and experimental immunology 2009;156:479-87.

33. Chen C, Coats SR, Bumgarner RE, et al. Hierarchical gene expression profiles of HUVEC stimulated by different lipid A structures obtained from Porphyromonas gingivalis and Escherichia coli. Cellular microbiology 2007;9:1028-38.

34. Nile CJ, Barksby E, Jitprasertwong P, et al. Expression and regulation of interleukin-33 in human monocytes. Immunology 2010;130:172-80.

35. Sanada S, Hakuno D, Higgins LJ, et al. IL-33 and ST2 comprise a critical biomechanically induced and cardioprotective signaling system. The Journal of clinical investigation 2007;117:1538-49.

[Downloaded from ijmcmed.org on 2026-09-16]