Investigation of LRTOMT Gene (Locus DFNB63) Mutations in Iranian Patients with Autosomal Recessive Non-Syndromic Hearing Loss

Seyyed Hossein Taghizadeh¹, Seyyed Reza Kazeminezhad¹, Seyyed Ali Asghar Sefidgar², Nasrin Yazdanpanahi³, Mohammad Amin Tabatabaeifar⁴, Ahmad Yousefi⁵, Seyyed Mohammad Lesani⁶, Marziyeh Abolhasani⁷, Morteza Hashemzadeh Chaleshtori^{7*}

- 1. Department of Genetic, Faculty of Science, ShahidChamran University of Ahwaz, Ahwaz, Iran.
- 2. Cellular and Molecular Biology Research Center (CMBRC), Babol University of Medical Sciences, Babol, Iran.
- 3. Department of Biochemistry and Genetics, Falavarjan Branch, Islamic Azad University, Isfahan, Iran.
- 4- Department of Genetics, Faculty of Medicine, JundiShapur University of Medical Sciences, Ahwaz, Iran.
- 5. Department of Basic Sciences, Faculty of Veterinary Medicine, Shahrekord University, Shahrekord, Iran.
- 6. Department of Genetics, Faculty of Science, Shahrekord University, Shahrekord, Iran.
- 7. Cellular and Molecular Research Center, Shahrekord University of Medical Sciences, Shahrekord, Iran.

Submmited 9 February 2013; Accepted 3 March 2013

Hearing loss (HL) is the most frequent sensory defect affecting 1 in 1000 neonates. This can occur due to genetic or environmental causes or both. The genetic causes are very heterogenous and over 100 loci have been identified to cause autosomal recessive non - syndromic hearing loss (ARNSHL). The aim of this study was to determine the contribution of the LRTOMT gene mutations in causing ARNSHL. One hundred fifty seven pupils affected with ARNSHL from Azarbaijan Sharghi, Kordestan, Gilan and Golestan provinces, north and west of Iran, were ascertained. In this descriptive - laboratory study, the presence of *LRTOMT* mutations were initially checked using PCR – Single - strand conformation polymorphism (SSCP) and heteroduplex analysis (HA) strategy. Samples with shifted bands on the gel were confirmed by DNA sequencing method. The PCR-SSCP/HA and the subsequent direct DNA sequencing showed no mutation in the population studied. We conclude that *LRTOMT* mutations have no role in causing sporadic deafness in the studied population. Further studies on other populations and samples could clarify the exact role of *LRTOMT* mutations.

Key words: Non - syndromic sporadic hearing loss, DFNB63, LRTOMT gene, PCR - SSCP, heteroduplex, Iran

Hearing loss (HL) is the most common sensory defect in human. One infant out of 1000 has pre - lingual HL. Some degrees of HL affect 4% of

the people under 45 and 10% of 65 or above (1-3). HL has a wide spectrum of clinical manifestations: congenital or late onset, conductional or sensory -

Corresponding author: Cellular and Molecular Research Center, Shahrekord University of Medical Sciences, Shahrekord, Iran. E-mail: mchalesh@yahoo.com.

nervous, syndromic or non - syndromic (4). The causes of HL can be genetical, environmental or both. More than 60% of the cases are assumed to be genetic. Over 70% of the genetic HL, are non - syndromic in which HL is the only evident manifestation. Almost 80% of the latter are autosomal recessive (1, 3-4). The phenotype is generally more severe in autosomal recessive non - syndromic HL (ARNSHL) and is usually manifested in pre-lingual cases (4, 5).

The estimations show that 1% of the human genes are involved in the hearing process. More than 100 loci have been identified in NSHL. Thus, HL could be regarded as one of the most heterogeneous traits in human (2).

Thus far, few studies have been performed on the sporadic form of NSHL in Iran and most of the investigation have been limited to a specific locus, DFNB1 (connexin-26 gene). Genetic studies of other loci are necessary to identify the causes and possible mechanisms of this disease. The results of these studies can definitely help to screen this type of HL and perform a more suitable genetic counseling.

To date, few investigations have been carried out about LRTOMT mutations in Iran and frequency of these mutations is not exactly known among Iranian ARNSHL patients (6). LRTOMT gene consists of 10 exons with 2 alternate reading frames and encodes two different proteins: LRTOMT 1 and LRTOMT 2 which are different in starting the translating codons. Exons 5, 7 and 8 have double reading frame. LRTOMT 1 has a transmembrane protein enriched of leucine whose function is unclear. LRTOMT 2 is an O - methyl transferase. The defects in O – methyl transferase protein have been noted to cause NSHL (7).

The current study was carried out on HL patients recruited from different provinces of Iran to screen exons 1, 2, 3, 5 and 8 of the *LRTOMT* gene (DFNB 63 locus) for mutations.

Materials and Methods

Sampling

One hundred fifty seven unrelated Iranian ARNSHL pupils from 13 elementary schools with average age of 11 years from four provinces of north and west of Iran (Azarbaijan Sharghi, Kordestan, Gilan and Golestan provinces) were included in this descriptive - laboratory study in 2011. About 5 ml peripheral blood sample from each individual was collected in tubes containing EDTA (0.5 ml). The samples were kept in - 20°C until use. A questionnaire was filled out for any family detailing clinical and demographical information and drawing family pedigree. All patients had mild to severe HL, were negative for GJB 2 mutations in our previous study and had not syndromic deafness, inflammatory middle ear disease or specific environmental ethiology according to examination performed by audiologist and general physician (8).

The study was approved by the Review Board of Shahrekord University of Medical Sciences.

DNA extraction

Salting out and phenol-chloroform methods were exploited for DNA extraction.

PCR: DNA samples were amplified using PCR method. Primers were designed and then blasted using the Gene Runner software (Hastings Software Inc, Hastings, NY). For exon 5, two sets of overlapping primers were designed due to its lengthy size (Table1). Site Directed Mutagenesis method by PCR, using primers with changes in one nucleotide, was used to make positive control samples. Standard PCR optimization was applied separately for each amplicon in 25 μ l volume reactions in a thermal cycler machine.

SSCP and HA

Microtubes containing PCR products were put in 96°C for 15 minutes. The samples were transferred on the ice immediately and kept in ice before loading on the polyacrylamide gel. TBE

Investigation of LRTOMT Gene (Locus DFNB63) Mutations

buffer, 1x was used in the gel and TBE 0.6 x was used in the tank (Table 2). Both mutated samples and hetero-duplex analysis were used to increase the sensitivity of SSCP. In order to perform hetero-duplex analysis, 3 ml EDTA (0.5M) was added to 2 ml PCR products for each sample. Bands were visualized by standard silver staining (9).

Sequencing of PCR products

The suspected samples with shifts on the SSCP gel and hetero-duplex were subjected to direct DNA sequencing of exons 1, 2, 3, 5 and 8 of

the LRTOMT gene in an ABI 3730XL automated sequencer (Applied Biosystems) (Macrogen, South Korea) for the final verification.

Results

A total of 157 deaf students, consisting of 85 males and 72 females age ranges from 9 to 13, from families with a history of ARNSHL, who suffered from mild to profound sensory neural HL with no other clinical findings were recruited. After applying quality control measures, no shifts were seen for the tested exons.

Exon	Pieces size (bp)	Primer Type	Primer sequence $(3 \rightarrow 5)$		
		F	GAAAGCAGTTGCCATGGAGT		
E1	196	R	GTGGGGGAAATCTCAGATCC		
		F	CAGCTTTACTTTAGCTAACAAATTGGA		
E2	290	R	AGCCCCATTACCTTTCCATC		
E3a	248	F	GCAAGGATGCAAGGAAGAGT		
		R	GCTCCTGTACCGAAGTGTTC		
		*RM	GCTCCTGTACCGAAGGGTTC		
E3b		F	GCACCTTTGAATGATGGCCT		
	259	R	ACTCTCCCCTACCCTCCAAA		
		FM*	GCACCTTTGAATGATTGCCT		
E5a		F	AGTATGGCTGTGGAGGGTTG		
	270	R	TTCCTCCATGGGGTTCCCAT		
		RM*	TTCCTCCATGGGGTTACCAT		
E5b		F	GTGAATAAGCTGGCTGTCCT		
	277	R	CAAGGAATGGAGGGACTTGA		
		FM*	GTGAATAAGCTGGCGGTCCT		
E8a		F	AGGATAATAATTGCTACTGGCAAAA		
	264	R	GTGAGCACGTAGCTGAAG		
		RM*	GTGAGCACGTAGCGGAAG		
E8b		F	GGCACTACTTCCGATTGCTG		
	262	R	ATCCCAAATATTCCTTCACTGTCTT		
		FM*	GGCACTACTTCCGATGGCTG		

M*: Mutant primers

Table 2: SSCP condition for exons of LRTOMT gene								
Exon	Gel density(%)	Time(h)	MA	Voltage(V)	Temperature(°C)			
Exon1	8	6.5	32	280	10			
Exon2	10	14	27	200	10			
Exon3a	8	7	31	300	10			
Exon3b	8	6.5	29	200	10			
Exon5a	8	7.5	32	200	10			
Exon5b	8	7.5	31	200	10			
Exon8a	8	7.5	32	280	10			
Exon8b	8	8	30	260	10			

Discussion

As all subjects in this study had not GJB2 mutations, the most prevalent deafness responsible gene, in our previous study (8) and the frequency of LRTOMT mutations is not known exactly among Iranian deaf individuals, we decided to investigate these mutations among deaf students from Azerbaijan Sharghi, Kordestan, Gilan and Golestan provinces, north and west of Iran.

Exons 1-5 and 8, including hot spot exons 2, 3, 8 of LRTOMT were selected for mutation analysis. The results showed the lack of mutation in the 157 studied samples negative for Connexin-26 gene mutations.

Overall, few studies have been done on this gene: Tlili et al. identified the DFNB63 locus on 11q13.3q13.4 chromosome related to deafness (1). Ahmed et al. identified the gene and the encoded protein structure. They reported 4 different homozygous mutations in *LRTOMT* gene (7). Finally, Du et al. analyzed the *LRTOMT* gene in 192 deaf children and identified homozygosity for a nonsense mutation in one family (11). On a series of Iranian autosomal recessive NSHL samples, a family was found to be affected with a 1 bp deletion (a frameshift mutation) (11). In the current study, the applied method was PCR-SSCP accompanied by HA, which was a simple, cheap and sensitive method (9). Using this combination, the sensitivity will increase considerably. In this study, all the variations introduced by site directed mutagenesis were recognized using the method.

Although other coding regions of the gene as well as the promoter of *LRTOMT* which were not studied in the current investigation, may be mutated, our findings suggest that the gene might play no considerable role in causing ARNSHL. However, further research is warranted for exact conclusion. Other exons and intronic sequences of LRTOMT should be investigated to clarify the role of this gene in causing deafness in Iranians.

Acknowledgement

We would like to sincerely thank all the students and their families for participating in this study. The contributions of the staff of the Exceptional Education and Training and Welfare Organizations of Azerbaijan Sharghi, Kordestan, Gilan and Golestan provinces are hereby apprecia-ted. This study was funded through a grant endowed by Shahrekord University of Medical Sciences, Iran.

Investigation of LRTOMT Gene (Locus DFNB63) Mutations References

 Gorlin RJ, Toriello HV, Cohen MM. Hereditary hearing loss and its syndromes. New York: Oxford University Press; 1995.
Petit C. Genes responsible for human hereditary deafness: symphony of a thousand. Nat Genet. 1996 Dec;14(4):385-91.

Morton NE. Genetic epidemiology of hearing impairment.
Ann N Y Acad Sci. 1991;630:16-31.

4. Marazita ML, Ploughman LM, Rawlings B, Remington E, Arnos KS, Nance WE. Genetic epidemiological studies of earlyonset deafness in the U.S. school-age population. Am J Med Genet. 1993 Jun 15;46(5):486-91.

5. Cohen MM, Gorlin RJ. Epidemiology, etiology and genetic patterns. In: Gorlin RJ, Toriello, H.V., Cohen, M.M., editor. Hereditary hearing loss and its syndromes. New York: Oxford University Press; 1995. p. 9-21.

6. Babanejad M, Fattahi Z, Bazazzadegan N, Nishimura C, Meyer N, Nikzat N, et al. A comprehensive study to determine heterogeneity of autosomal recessive nonsyndromic hearing loss in Iran. Am J Med Genet A. 2012 Oct;158A(10):2485-92.

7. Ahmed ZM, Masmoudi S, Kalay E, Belyantseva IA, Mosrati

MA, Collin RWJ, et al. Mutations of LRTOMT, a fusion gene with alternative reading frames, cause nonsyndromic deafness in humans. Nat Genet. 2008 Nov;40(11):1335-40.

8. Chaleshtori MH, Farhud DD, Patton MA. Familial and sporadic GJB2-related deafness in iran: Review of gene mutations. Iran J Public Health. 2007;36(1):1-14.

9. Sunnucks P, Wilson ACC, Beheregaray LB, Zenger K, French J, Taylor AC. SSCP is not so difficult: the application and utility of single-stranded conformation polymorphism in evolutionary biology and molecular ecology. Mol Ecol. 2000 Nov;9(11):1699-710.

10. Tlili A, Masmoudi S, Dhouib H, Bouaziz S, Rebeh IB, Chouchen J, et al. Localization of a novel autosomal recessive non-syndromic hearing impairment locus DFNB63 to chromosome 11q13.3-q13.4. Ann Hum Genet. 2007 Mar;71(Pt 2):271-5.

11. Du X, Schwander M, Moresco EMY, Viviani P, Haller C, Hildebrand MS, et al. A catechol-O-methyltransferase that is essential for auditory function in mice and humans. P Natl Acad Sci USA. 2008 Sep 23;105(38):14609-14.