A comparison between cytotoxicity induced by two resin based sealers (2Seal and AH Plus) in Saos-2 and MG-63 cell lines

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The aim of this study was to evaluate and compare the cytotoxicity induced by two resin-based sealers, 2Seal and AH Plus, in two osteoblast-like cell lines, MG-63 and Saos-2. Using sterile discs of both sealers in complete media, 24- and 72-h extracts were prepared. The extracts were exchanged with Saos-2 or MG-63 cell culture media at 75% confluence, and after 24 h incubation, cell viability tests were performed for each extract and cell line using MTT and trypan blue dye exclusion assays. Corresponding incubated media were used as negative control groups. For both extracts and sealers, cytotoxicity was observed in both cell lines. For Saos-2, there was no statistical difference in toxicity between the sealers for either extract (p > 0.05). For MG-63, the 2Seal 24-h extract and the AH Plus 72-h extract had greater cytotoxicity than the other extracts (p < 0.05). Both AH Plus and 2Seal demonstrated significant cytotoxicity in these two cell lines. In contrast to 2Seal, the cytotoxicity of AH Plus in the MG-63 cell line increased with extraction time from 24 to 72 h. The AH Plus and 2Seal 24-h extracts showed different levels of cytotoxicity in the MG-63 cell line, while in the Saos-2 cell line there were no detectable differences. This may reflect higher sensitivity of the MG-63 cell line compared to Saos-2 toward cytotoxicity induced by these two sealers, or different kinetics of toxicant release from the sealers.

Key words: Cytotoxicity, AH Plus, 2Seal, osteosarcoma, Saos 2, MG-63

Substances used for root canal sealing along with endodontics treatment procedures have improved considerably during the last two centuries (1). Despite of great achievements in this field, investigations are ongoing toward developing materials with better physico-chemical properties and lower toxicities. The ideal root canal sealer should prevent penetration of periapical exudates into root canal, prevent recurrence of infection, and provide a microenvironment suitable for tissue healing (2, 3). On the other hand, biocompatibility of the root canal sealers, which could be directly or indirectly in contact with periapical tissues, is very important (4-6) and could affect the healing

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Cell culture based cytotoxicity assays for medical devices and dental materials have got great approvals in the recent decades compared to exhaustive and time consuming in vivo models (4, 7).

Numerous cell lines including those obtained from human periodontal fibroblasts have been used for dental materials cytotoxicity assays (8, 9). Also cell lines originated from tissues other than periapical or human oral cavity (e.g. 3T3, Hela, V79…) (7, 10, 11) have been used in these assays. However, in order to have a better prediction on biocompatibility of tested compound, it is preferred to use cell lines with similar characteristics and phenotypes to dental and periapical tissues (12, 13). Since osteoblasts play an important role in healing dental and apical tissues, we chose two osteosarcoma cell lines with human origin “Saos-2 and MG-63” (14-16).

In view of the fact that the chemical composition of different sealing materials varies from one type to another, the in vitro biocompatibility results would depend on the selected method of cytotoxicity assay (6). In addition to duration of extraction, the type of cell line, and exposure method would also affect the in vitro biocompatibility results. Some materials do not release toxic substances so much but show deleterious effects when come to contact with cells or tissues. At these cases, putting set discs directly in the culture vessels would simulate the in vivo condition and more likely detect cytotoxic effects. Gutta-percha is one of the most common used root canal filling materials so far which has a very good biocompatibility (7) but other sealing materials such as zinc oxide-eugenol cement, calcium hydroxide-based, and resin-based sealers release toxic substances and show different degrees of cytotoxicities (5, 9). AH Plus as a well known epoxy resin-based sealer, has shown good properties for successful endodontic therapy including less formaldehyde release, hence lower cytotoxicity in many cell lines (8, 17, 18). As a relatively new introduced sealing material, 2Seal has many physico-chemical properties in common with AH Plus, including its epoxy-amine resin based composition and according to the manufacturer it does have minimal toxicity on living tissues (19). In this study we aimed to evaluate the cytotoxicity induced by 2Seal and compare it with its common used congener AH Plus on two osteosarcoma (fibroblast like) cell lines, Saos-2 and MG-63.

### Methods and Materials

#### Cell culture condition

Saos-2 and MG-63 cell lines (Pasteur Institute, Iran) were seeded in 24 well plates (1.5×10⁵ cells/well) with RPMI-1640 (Sigma-Aldrich, UK) supplemented with 10% FBS (Invitrogen, UK) and 1% PenStrep® (Sigma-Aldrich, UK). After 24 hours, when cells reached 70-80% confluence, the media were changed by the corresponding sealers extracts and after another 24 hours the viability of the cells were measured.

#### Preparation and extraction of the sealers

According to the manufacturers’ instruction, components of either AH Plus (DENTSPLY DETREY GmbH, Germany) or 2Seal (Roydent Dental Products, USA) were mixed and cylindrical discs (3 mm diameter and 3 mm thickness) were made in a laminar flow safety cabinet using a stainless steel mould. Then discs were aseptically incubated at 37°C for 24 hrs to be completely set. Each disc was drawn in 1 ml complete media (RPMI-1640+10% FBS+1% PenStrep®) in 24 wells cell culture plates. The supernatants were collected after 24 or 72 hrs and replaced with either MG-63 or Saos-2 culture media. Complete media incubated in empty wells was used as negative control.

#### Sealers cytotoxicity assays

All experiments were performed in triplicates. The viability of the cells was measured after Saos-2 or MG-63 cell lines exposure to the
A comparison between cytotoxicity induced by two resin sealers extracts using MTT assay (20) and Trypan blue dye exclusion (TB) method (21). Briefly, after washing the cells with D-PBSA, 200μl of MTT (Sigma-Aldrich, UK) solution in PBS (5 mg/ml) was added to each well and incubated for 4 hrs at 37°C. The purple/blue formazan precipitate was dissolved in 800 μl of acidic isopropyl alcohol (0.04 N) and the colored solution absorbance was read at 570 nm and 630 nm (as reference wavelength) using a UV-Vis spectrophotometer. For TB assay, numbers of viable cells after 24 hours exposure to the extracts were counted for each triplicate and presented as percentage of the control.

**Statistical Analysis**

The results have been presented as Mean ± Standard Error. Different groups’ means were compared by t-Student and one-way analysis of variance (ANOVA) and Tukey’s Post Hoc tests. The statistical significance was set at p< 0.05.

The results of MTT assays for both 24 and 72 hrs extracts show significant cytotoxic effects in MG-63 and Saos-2 cell lines induced by 2Seal and AH Plus (Fig. 1 and 2). There were no statistically differences between 2Seal and AH Plus cytotoxicities in Saos-2 cell line with both 24 and 72 hrs extracts (Fig. 1). However, there was a higher cytotoxicity on MG-63 observed by 24 hrs extract of 2Seal which decreased in favor of AH Plus with 72 hrs extract (Fig. 2). Results obtained by trypan blue assays showed similar pattern of decrease in cell numbers (Fig. 3, 4).

**Discussion**

Evaluation of cytotoxicity induced by root canal filling materials using human cell lines has been widely used in recent decades to predict and compare the new materials biocompatibilities (4, 6). In the current study we compared cytotoxicity of two common used root canal sealers (AH Plus and 2Seal) on 2 fibroblast like cell lines (Saos-2 and MG-63). Based on previous investigations conducted worldwide, AH Plus had showed prior biocompatibility to many other resin based sealers (11, 17, 22, 23). Our results show that both sealers with 24 or 72 hrs h extraction times have the same cytotoxic effects on Saos-2 cell line, however on MG-63 cell line, 2Seal shows more cytotoxicity with 24 hrs extract while AH Plus is more toxic after 72 h extraction.
Despite of worldwide AH Plus’ common use, it causes cytotoxic effects on different cell lines, like other root canal filling materials (e.g. on murine fibroblast, V79, Hela...) (8, 10, 17, 18, 24, 25). The in vitro method chosen to test the cytotoxicity of root filling materials could significantly influence the obtained final results. Freshly made mixtures usually show more cytotoxic properties than their set forms (11, 18, 24). These mixtures might disintegrate in the liquid media used in the cell culture and do not necessarily resemble the normal in vivo condition existing in human dental canal. Using set discs of AH Plus and 2Seal with same size and shape keeps the extraction ratio of surface area per extraction volume constant, hence elute concentration would be only dependent on the nature of the dissolved substances.

It has been shown that with different sealer materials, extraction time might have great effects on the final observed cytotoxicities. For instance with 24 hrs extraction time, Endion has showed higher toxicity on Saos-2 cell line compared to AH Plus, while with 72 hrs extraction time, AH Plus toxicity exceeded (26). AH Plus is an epoxy resin-based sealer which seems to release more toxic substances into the medium during 72 hrs incubation, hence its cytotoxicity increases by time compared to a silicone-based sealer (e.g. Roeko Seal Automix) (17). We found similar results with MG-63 cell line, where AH Plus showed higher cytotoxicity with 72 hrs extract compared to 2Seal that showed higher toxicity with 24 hrs extract (Fig. 2, 4). However, such a difference was not statistically significant in Saos-2 cell line which might be due to the difference in sensitivity of these two cell lines to the eluted toxic compounds (12) and/or the instability of eluted toxic substances in the media. The instability could be due to the fact that formaldehyde and other volatile species might leave the media by warm incubation (22).

Alternatively some reactive species might cross react with the serum proteins in the media with consequent decrease in toxicity during extraction time. Since AH Plus setting process is the result of a polymerization system called “Linear Epoxide-Amine Addition”, leaching non reacted monomers from this polymer matrix, might be another justification for its constant increasing (however less than the other sealing materials) cytotoxicity with longer extraction time (4, 27, 28).

On the other hand, there is little information concerning 2Seal cytotoxicity on different cell lines especially Saos-2 and MG-63 cell lines. The information provided by the manufacturer indicates
A comparison between cytotoxicity induced by two resin that 2Seal (like AH Plus) is a polymer made from interaction between a bisphenol derivative with a diamine derivative. It is an epoxy resin sealer and leftover of both these two copolymers are toxic and might be responsible for 2Seal cytotoxicity. In an in vivo study conducted with 3 different sealers on rat molars, 2Seal showed the best biocompatibility in terms of producing less periapical inflammation compared to the other two sealers: RSA Roekoseal and Aptal Harz (29).

In another study, 2Seal had showed histological effects similar to AH Plus on canine molars periapical tissues (23). In the present study, at least on Saos-2 cell line, both sealers showed similar toxicities. Different sensitivities to sealing materials elutes observed in different cell lines has been reported in many other studies (12, 21, 30) and it has been recommended to perform cytotoxicity studies on different cell lines before any discrete judgment about biocompatibility of different biomaterials (12). The difference between kinetic of toxic substances release from solidified polymers might be the reason for slight differences observed by different extraction times with MG-63 cell line. However, the difference between each cell line susceptibility to the type of elutes should not be ignored as well.

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References